

# **Review and Selection of Fish Metrics**

## **for the Chicago Area Waterway System**

### **Habitat Evaluation and Improvement Study**

**Prepared for**  
**The Metropolitan Water Reclamation District**  
**Of Greater Chicago**

April 21, 2009

**This page is blank to facilitate double sided printing.**

## TABLE OF CONTENTS

1. Introduction.....	1
2. Data Description .....	3
2.1 Fish Sampling Locations.....	3
2.2 Fish Sampling Methods .....	3
2.3 Summary Description of Fish Data.....	4
3. Selection of Fish Metrics .....	19
3.1 Compilation of Fish Metrics .....	19
3.2 Specification of Tolerance Values .....	22
4. Screening of Fish Metrics .....	25
4.1 Screening Objectives .....	25
4.2 Metrics Lacking Data.....	25
4.3 Metric Range.....	25
4.4 Metric Redundancy .....	26
4.5 Metric Variability.....	27
5. Final recommended List of Metrics .....	33
6. References.....	35

**LIST OF FIGURES**

Figure 2-1. Fish Sampling Stations in the CAWS.....	5
Figure 2-2. Taxa Collected among CAWS Stations for the 2001-2007 sample period. Blue bars indicate stations included in the quadrennial sampling schedule while the orange bars indicate those sampled annually.....	6
Figure 2-3. Taxonomic Abundances across the CAWS at Annual Monitoring Stations.....	8
Figure 2-4. Species Observations, by Sample Event at all Monitoring Stations for the 2001-2007 Period.....	10
Figure 2-5. Species Observations, by Sample Event at Annual Monitoring Station for the 2001-2007 Period.....	11
Figure 2-6. Total Number of Individuals Collected during the 2001-2007 Sample Period (black bars are referenced to the axis on the left, blue bars are referenced to the axis on the right). ....	12
Figure 2-7. 2001 to 2007 Annual Station Fish Survey Results .....	14
Figure 2-8. Results of Electrofishing and Fyke Net Samples by Length Interval , from 2007 Samples near Harlem Avenue on the Cal-Sag Channel. ....	16
Figure 2-9. Results of Electrofishing and Fyke Net Samples by Length Interval , from 2007 Samples near Southwest Highway on the Cal-Sag Channel. ....	17
Figure 2-10. Results of Electrofishing and Fyke Net Samples by Length Interval , from 2007 Samples near Cicero Avenue on the Cal-Sag Channel.....	18
Figure 4-1. Coefficient of Variation for %LTHPL_(wt) and %LTHPL_(n), for 2001 through 2007 Data. ....	28
Figure 4-2. Coefficient of Variation for %LTHPL_(wt) and %LTHPL_(n) at Annual Sampling Stations. ....	29
Figure 4-3. Coefficient of Variation for %INSCT_(wt) and % INSCT_(n), for 2001 through 2007 Data. ....	29
Figure 4-4. Coefficient of Variation for %INSCT_(wt) and % INSCT_(n) at Annual Sampling Stations. ....	30
Figure 4-5. Coefficient of Variation for %TC_(wt) and % TC_(n), for 2001 through 2007 Data. ....	31
Figure 4-6. Coefficient of Variation for %TC_(wt) and % TC_(n) at Annual Sampling Stations. ....	31
Figure 4-7. Coefficient of Variation for %INT_(n) and INTOL, for 2001 through 2007 Data.....	32
Figure 4-8. Coefficient of Variation for %INT_(n) and INTOL at Annual Sampling Stations. ....	32

**LIST OF TABLES**

Table 2-1. Taxa Richness and Total Number of Individuals by Station and Year. ....	7
Table 3-1. Initial List of Fish Metrics.....	21
Table 4-1. Fish Metrics Remaining after Screening for Redundancy. ....	27
Table 5-1. Final Recommended Fish Metrics for Use in the CAWS. ....	34

## **LIST OF ATTACHMENTS**

Attachment A: CAWS Fish Data Stations and Sampling Dates Used in this Analysis

Attachment B: List of Fish Species Identified in the CAWS (2001-2007) and Their  
Tolerance Assignments

Attachment C: Matrix of Pearson's Correlation Values for Fish Metrics

**This page is blank to facilitate double sided printing.**

## 1. INTRODUCTION

This document summarizes the process used to select key fish metrics for the Chicago Area Waterway System (CAWS) Habitat Evaluation and Improvement Study (the Study). Selection of key fish metrics is important to the Study for two reasons:

- **Sensitivity to Water Quality** – Comparison of historical fish data and water quality data is important in understanding the relationship between water quality and fish communities in the CAWS. Identification of CAWS appropriate fish metrics is necessary for such data comparisons.
- **Habitat Index Development** – The proposed method for development of a CAWS-specific habitat index relies on the comparison of fish data to habitat variables to help define the relationship between fish and the physical habitat in the CAWS.

It was not the objective of the Study to develop a CAWS-specific index of biotic integrity (IBI), but the methods used to identify key fish metrics for the CAWS are the same as those used in current biological practice to define metrics for fish IBIs. Development of a fish IBI for the CAWS might be useful in the future, but development an IBI would require specification of a regionally appropriate, non-consumption, target condition to which the upper end of the index would be referenced (Karr 1991). This can be done in one of three ways, but is currently beyond the scope of this analysis for the CAWS as described below:

- **External reference reach** – An external reference reach that represents a target fisheries condition that is attainable in the CAWS could be used to establish the upper limit of the IBI. This approach is impractical for the CAWS because the CAWS consists entirely of constructed or heavily modified channels and no similar channels with high quality or reference fisheries have been identified.
- **Internal reference reach** – A reach within the system that represents a target fisheries condition that should be targeted for the entire CAWS could be used to establish the upper limit of the IBI. This is not currently possible because no such internal reference has been identified.
- **Target use** – A target fisheries use (e.g., warm water sport fishing), function (e.g., harvest prohibition) or specific target species (e.g., trophy largemouth bass) may be identified which would allow determination of target fisheries conditions to describe the upper end of the index. To date, target uses or species have not been identified.

Although it is currently impractical to establish a fish IBI for the CAWS, it is possible to determine key fish metrics for use in comparing to habitat data. This document presents the recommended list of fish metrics for the CAWS and summarizes the methodology used to arrive at that list.

**This page is blank to facilitate double sided printing.**



## 2. DATA DESCRIPTION

This section provides an overview of the fish data used in this study.

### 2.1 FISH SAMPLING LOCATIONS

The Metropolitan Water Reclamation District of Greater Chicago (District) has been collecting fish data annually since 1974 (with the exception of 1981 and 1982) within the CAWS. However, to focus this Study on current conditions, LimnoTech limited the fish data analysis to the data collected between 2001 through 2007 and to the area considered as the managed portion of the CAWS. The managed portion is defined by the non-wadeable waters bounded by the Wilmette Pumping Station, the Chicago River Lock and Controlling Works, the O'Brien Lock and Controlling Works and the Lockport Lock and Powerhouse. The tributaries to the CAWS are not included in this study, as their physical conditions and regulatory controls differ from the mainstems of the CAWS. The South Fork of the South Branch, also known as Bubbly Creek, is also included in this study.

During the 2001-2007 period, the District collected fish data at 34 stations within the CAWS (Figure 2.1) on a routine basis. Twenty-six of these 34 stations are part of the District's Ambient Water Quality Monitoring (AWQM) program. Seven of the AWQM stations are annually monitored (once per year), while the remainder are sampled on a four year rotation. The total number of sample events across all stations and years includes 113 sample events. The CAWS fish monitoring stations and sampling dates used in the sample description, screening and selection of fish metrics is included as Attachment A.

### 2.2 FISH SAMPLING METHODS

The District samples the fishery within the CAWS using boat electrofishing procedures<sup>1</sup>, following standard and consistent protocols for this collection method. Each station is generally defined by a 400 meter reach and each bank length was sampled for fishes. The average shock time averages 800 seconds. The collected fish are counted, measured (standard and total length), weighed and released, except where difficult to identify in the field. In addition, any abnormalities such as diseases, eroded fins, lesions or tumors (DELTs) are noted. Between 2001 through 2007, all sampled stations have a single sampling event per year except Station 75, Chicago Sanitary and Ship Canal at Cicero Avenue. During the first sampling event on 7/31/2001 the field crew experienced equipment failure, which resulted in a partial fish collection sample. Later in the season, on 9/4/2001 the crew returned to the station to conduct an additional sampling. Only the 9/4/2001 data were included in this study. Finally, supplemental sampling was conducted in 2007 using Fyke nets at three stations, and those data are also summarized.

---

<sup>1</sup> In 2007, the District supplemented fish collections with Fyke net samples but, because this method is not consistent with other methods, these data were not included in this analysis.

## **2.3 SUMMARY DESCRIPTION OF FISH DATA**

Fifty-two (52) species, including five hybrids, of fish were identified at the 34 CAWS monitoring stations between 2001 and 2007 (sample period). Attachment B provides the complete list of these fish species. For the sample period, the number of non-hybrid species collected across the CAWS stations ranged from 27 at AWQM Station 76 (Little Calumet River at Halsted Street) to only five at Stephen Street (Chicago Sanitary Shipping Canal; CSSC; Figure 2-2). The repeated, annual sampling effort did not necessarily relate to the greatest number of taxa among the sample period for an individual station. For example, the second most numerous taxa (n=23), were from the Little Calumet River at Indiana Avenue, resulting from only two sample events for the sample period. Figure 2-1 depicts the distribution of the number of non-hybrid collected taxa across the managed portion of the CAWS. Table 2-1 describes the taxa richness and total number of individuals by station, for the sample period.



Figure 2-1. Fish Sampling Stations in the CAWS.

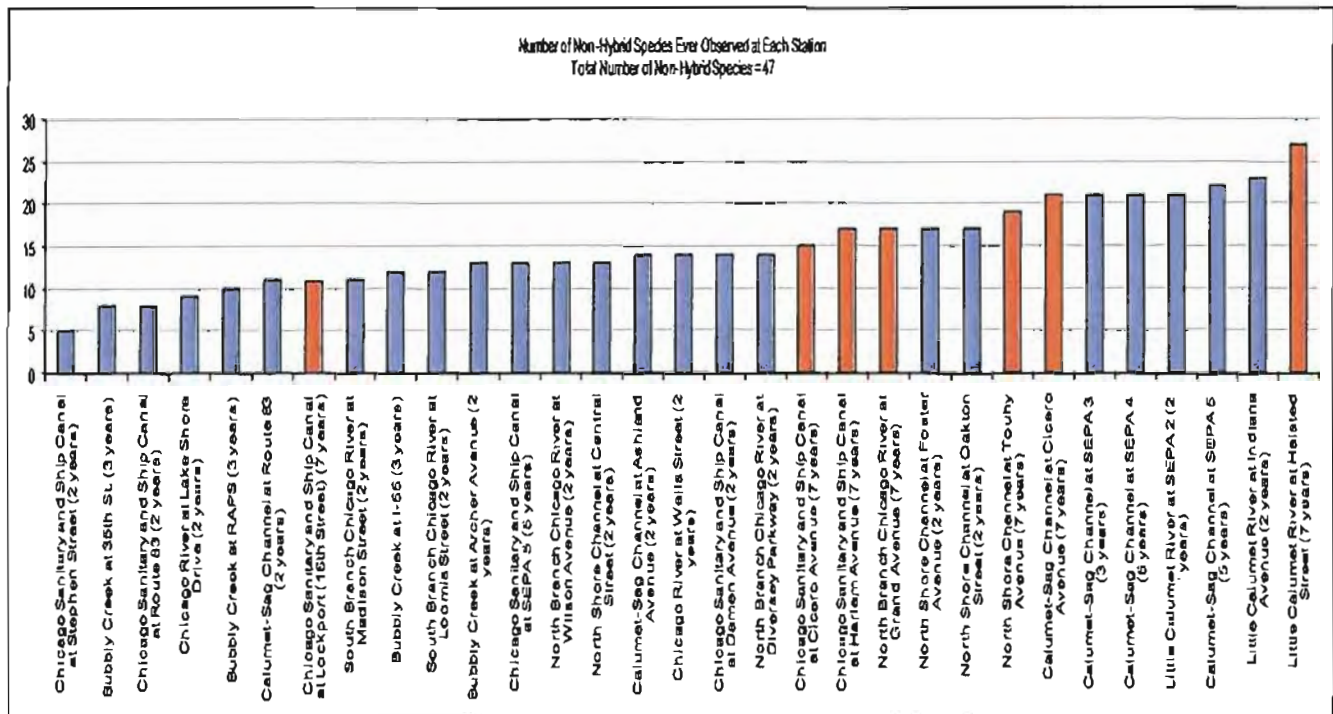


Figure 2-2. Taxa Collected among CAWS Stations for the 2001-2007 sample period. Blue bars indicate stations included in the quadrennial sampling schedule while the orange bars indicate those sampled annually.

Table 2-1 summarizes the station sample collections by station and year. Station sampling within the CAWS has ranged from as few as 12 stations (2001) to as many as 20 stations (2005) with an average of 16 stations sampled per year. Station samples vary in their taxa and total number of individuals both within stations among years, and among stations. The least number of species collected in any event occurred in 2001 at Lockport with only 2 taxa represented by 77 individuals. The greatest number of species for a single event included 22 taxa represented by 405 individuals collected on the Little Calumet River at Halsted Street in 2006.

**Table 2-1. Taxa Richness and Total Number of Individuals by Station and Year.**

AWQM Station Number	Station Description	Sample taxa richness (total number of individuals)						
		2001	2002	2003	2004	2005	2006	2007
35	North Shore Channel at Central Street	12 (132)				11 (139)		
36	North Shore Channel at Touhy Avenue	11 (595)	12 (147)	14 (335)	11 (249)	9 (276)	16 (496)	14 (387)
101	North Shore Channel at Foster Avenue	15 (179)				17 (273)		
102	North Shore Channel at Oakton Street	2 (2)				17 (151)		
37	North Branch Chicago River at Wilson Avenue	9 (75)				11 (122)		
73	North Branch Chicago River at Diversey Parkway	7 (58)				13 (164)		
56	Little Calumet River at Indiana Avenue			17 (452)				18 (322)
76	Little Calumet River at Halsted Street	16 (210)	17 (163)	13 (219)	17 (207)	19 (913)	22 (405)	23 (281)
SEPA2	Little Calumet River at SEPA 2					16 (529)	12 (218)	
43	Calumet-Sag Channel at Route 83			7 (43)				9 (261)
58	Calumet-Sag Channel at Ashland Avenue			13 (95)				12 (131)
59	Calumet-Sag Channel at Cicero Avenue	10 (127)	13 (174)	12 (56)	10 (147)	10 (453)	15 (214)	12 (297)
SEPA3	Calumet-Sag Channel at SEPA 3			13 (148)		16 (253)		14 (407)
SEPA4	Calumet-Sag Channel at SEPA 4			11 (93)	11 (82)	14 (653)	9 (79)	15 (417)
SEPA5	Calumet-Sag Channel at SEPA 5			12 (232)	7 (41)	16 (443)	7 (37)	17 (216)
Supplemental Survey	Calumet-Sag Channel at 104th Street							10 (92)
Supplemental Survey	Calumet-Sag Channel at Kedzie Avenue							8 (87)
Supplemental Survey	Calumet-Sag Channel at Southwest Highway							13 (127)
46	North Branch Chicago River at Grand Avenue	12 (53)	7 (28)	8 (67)	9 (88)	5 (77)	10 (158)	13 (117)
74	Chicago River at Lake Shore Drive		8 (22)				7 (83)	
301	Chicago River at Wells Street		11 (136)				10 (250)	
39	South Branch Chicago River at Madison Street		10 (138)				6 (99)	
40	Chicago Sanitary and Ship Canal at Damen Avenue		10 (148)				12 (164)	
99	Bubbly Creek at Archer Avenue		5 (21)				13 (156)	
108	South Branch Chicago River at Loomis Street		10 (76)				13 (142)	
99.2	Bubbly Creek at 35th St.			5 (39)	8 (27)	5 (26)		
99.1	Bubbly Creek at I-55			6 (31)	10 (60)	5 (31)		
99.3	Bubbly Creek at RAPS			7 (151)	10 (97)	5 (62)		
41	Chicago Sanitary and Ship Canal at Harlem Avenue	9 (88)	11 (188)	10 (225)	13 (193)	14 (758)	15 (388)	12 (282)
42	Chicago Sanitary and Ship Canal at Route 83		5 (32)				5 (10)	
48	Chicago Sanitary and Ship Canal at Stephen Street		4 (24)				5 (24)	
75	Chicago Sanitary and Ship Canal at Cicero Avenue**	10 (118)	10 (136)	9 (138)	13 (191)	7 (184)	11 (206)	13 (280)
92	Chicago Sanitary and Ship Canal at Lockport (16th Street)	2 (77)	6 (67)	7 (67)	4 (22)	9 (179)	8 (64)	6 (64)
SEPAS_CSSC	Chicago Sanitary and Ship Canal at SEPA 5			5 (18)	8 (53)	6 (306)	8 (34)	9 (178)

Figure 2-3 depicts the sample variation among years at the annual stations. The figure also includes the annual variation of species assigned to pollution tolerance categories of tolerant (to pollution), intolerant and moderately tolerant. A discussion of the categorical assignments for pollution tolerance is included later and tolerance assignments for individuals are included in Attachment B. In general, the number of taxa collected within the annual monitoring stations appears to be increasing since 2001. Tolerant species dominate all annual stations, followed by moderately tolerant species. Several stations have no intolerant species represented during any sample year, while others have a few.



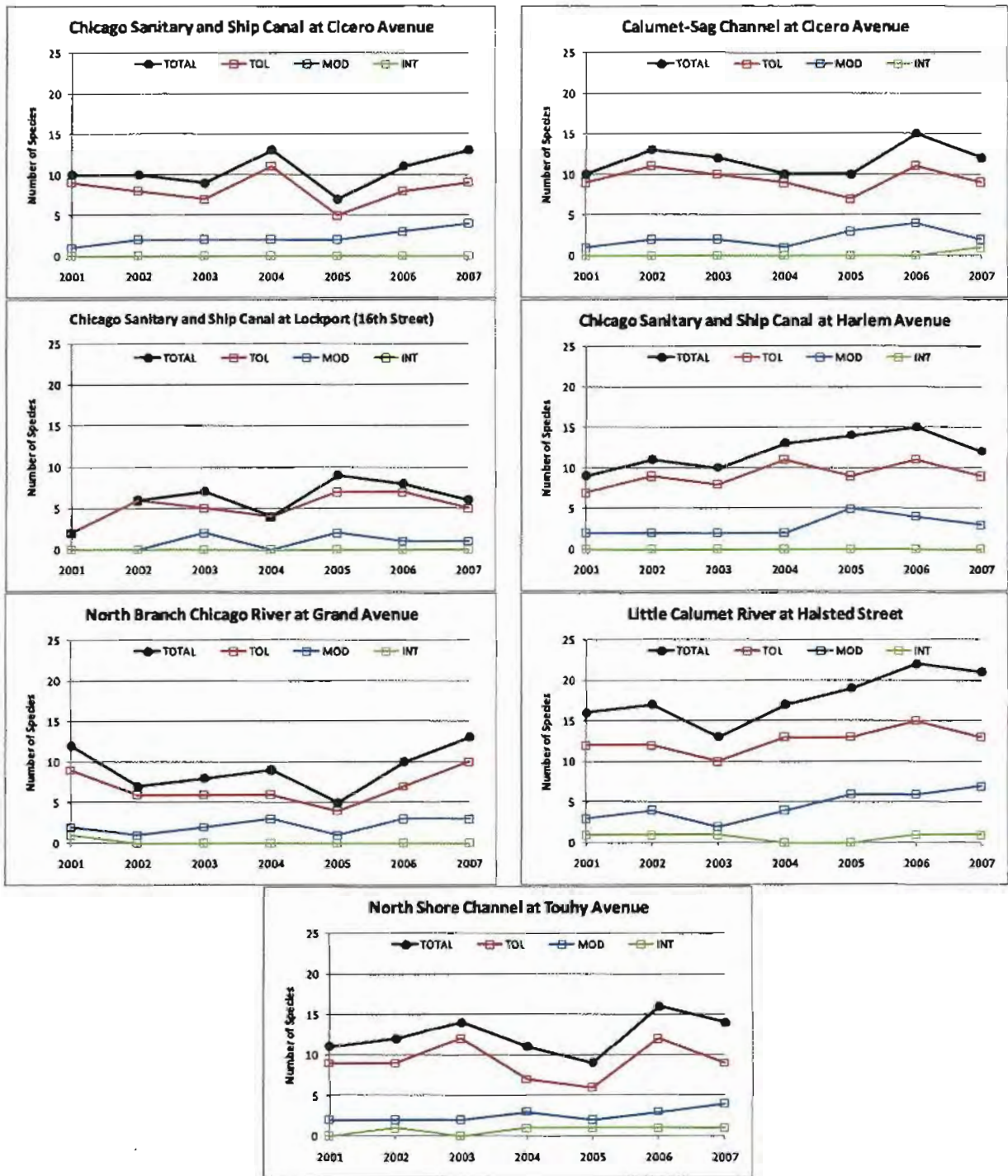


Figure 2-3. Taxonomic Abundances across the CAWS at Annual Monitoring Stations.

The most frequently observed species across all stations included gizzard shad (*Dorosoma cepedianum*), common carp (*Cyprinus carpio*), and largemouth bass (*Micropterus salmoides*), respectively (Figure 2-4). The most frequently observed species at the annual monitoring stations includes gizzard shad, common carp and pumpkinseed (*Lepomis gibbosus*), respectively (Figure 2-5). The most numerous observed species within the CAWS included gizzard shad (n=6906), emerald shiner (*Notropis atherinoides*; n=2082) and common carp (n= 2055), respectively (Figure 2-6). Eleven species are represented by only a single observation for the 2001-2007 period. Finally, gizzard shad, common carp, and largemouth bass have been observed at all stations during the sample period.

The distribution and abundance of gizzard shad in the CAWS is not unusual for large water systems and Simon and Sanders (1999) suggest not including this species in community structure comparisons as a potential source of bias in analysis. Emerald shiner is commonly found in large rivers and appears to thrive in reservoir systems (Becker 1983), so their numbers and distribution within the CAWS is not unexpected. Common carp are found turbid, warm, large river systems of the Midwest (Becker 1983) and their distribution and abundance in the CAWS is also not surprising. Largemouth bass are also abundant in large rivers of the Midwest (Becker 1983), with a presence expected in the CAWS and serve as a popular recreation target species within the system (Personal communication, Bradley 2008). Pumpkinseed also appears to thrive in impounded systems (Becker 1983) so their numbers and distributions are also not unexpected.

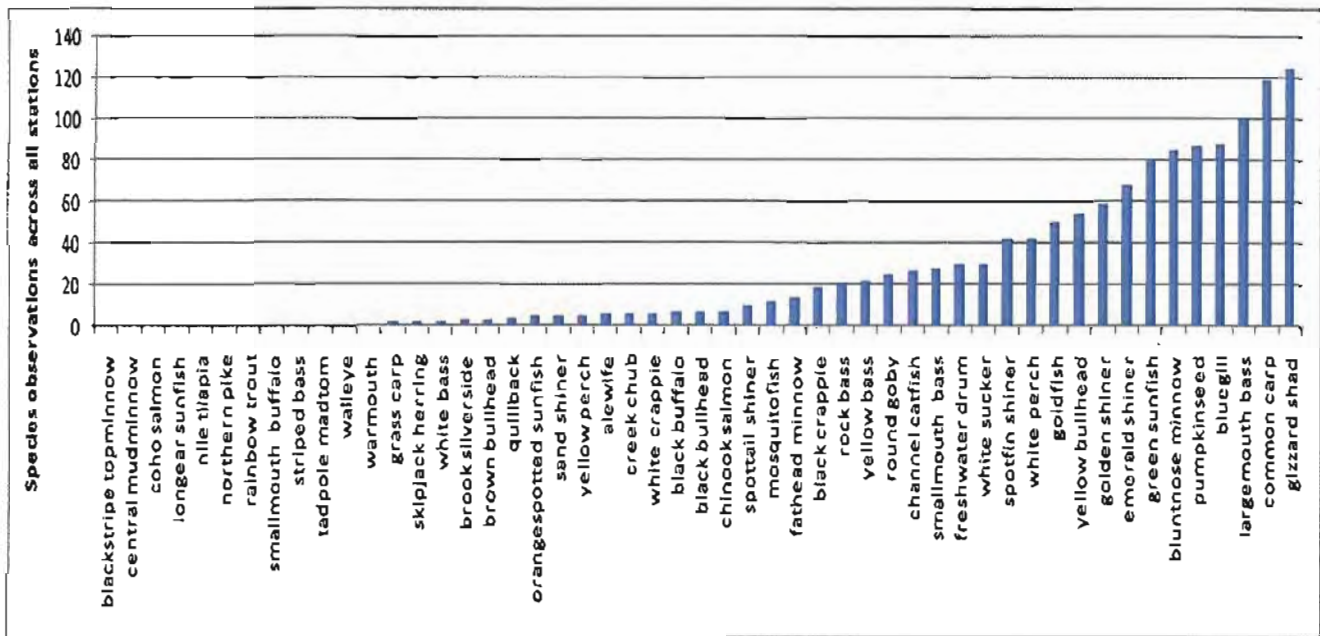


Figure 2-4. Species Observations, by Sample Event at all Monitoring Stations for the 2001-2007 Period.



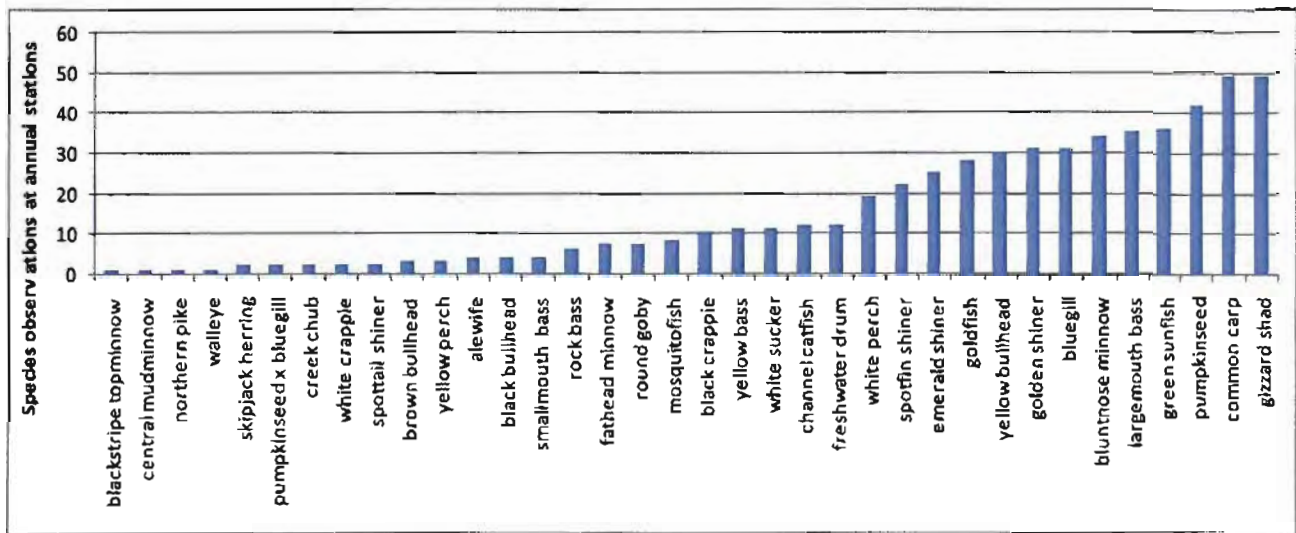


Figure 2-5. Species Observations, by Sample Event at Annual Monitoring Stations for the 2001-2007 Period.

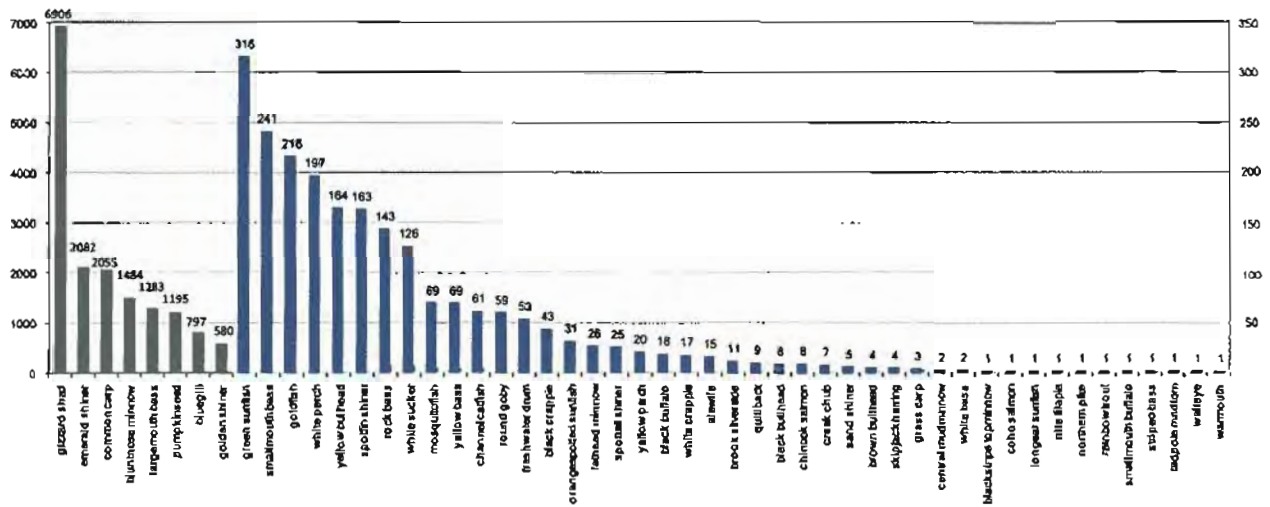


Figure 2-6. Total Number of Individuals Collected during the 2001-2007 Sample Period (black bars are referenced to the axis on the left, blue bars are referenced to the axis on the right).

Figure 2-7 describes the sample collections, among years at the annual monitoring stations. The graphs depict the variation of samples collected at the stations among years. Meador and McIntyre (2003) observed high variability (Coefficient of Variation (CV) = 0.23) in species richness from boat electrofished samples of up to +/-5 species and concluded that their variation resulted from sampling efficiency, rather than environmental variation. However, they noted that increased variation among years at a station was related to increasing station depth (Meador and McIntyre 2003). Paller (1995) suggests that a CV of 0.20 is the maximum desirable level of variability in catch per unit effort for electrofishing. The CV for the CAWS annual monitoring stations ranged from 0.16 – 0.4. The Lockport station had the highest CV (0.40), while the Cal-Sag station at Cicero Avenue had the lowest CV (0.16). The high CV at the Lockport station may be related to the site conditions of confined, deep channels, no access to shallow water areas and a species community that is dominated by mobile species such as gizzard shad, carp, and a range of sunfishes. These findings are also consistent with Meador and McIntyre (2003) in their descriptions of highly variable non-wadeable sites.

Finally, in 2007, the District deployed Fyke nets as a supplemental sampling method for three stations within the CAWS. The Fyke net collected data was compared to the closest electrofishing event in space and time in an attempt to understand how this additional collection method may be of value for use in the CAWS fishery monitoring program for capturing smaller age-class fish. Fyke nets are selective for migratory fish that follow shorelines (Hubert 1996). The 2007 samples resulted in relatively small catches compared to electrofishing and seemed biased towards smaller size classes (Figures 2-8, 2-9, 2-10). The Cal-Sag at Harlem Avenue resulted in the largest catch of 34 individuals. Of the 34 individuals, only four bluegill (total length ranging 31-37 mm) were collected with the remaining species being minnows. Only three individuals were collected at the Cal-Sag at Cicero Avenue site: two minnow and one bluegill (total length 31 mm). The Cal-Sag at Southwest Highway site found 11 individuals: 7 bluegill (total length 23-46 mm), one green sunfish (total length 48 mm), and the remaining were minnows. Overall, the catch total lengths from the Fyke net samples ranged between 23 mm and 66 mm. Little can be drawn from the small catches of the 2007 Fyke net sample data other than the samples seemed biased towards small samples of young, potentially year 1 (Becker 1983) bluegill and minnows. Future, alternative approaches may include light-traps that target young-of-year fishes to try to understand reproduction within various portions of the CAWS.

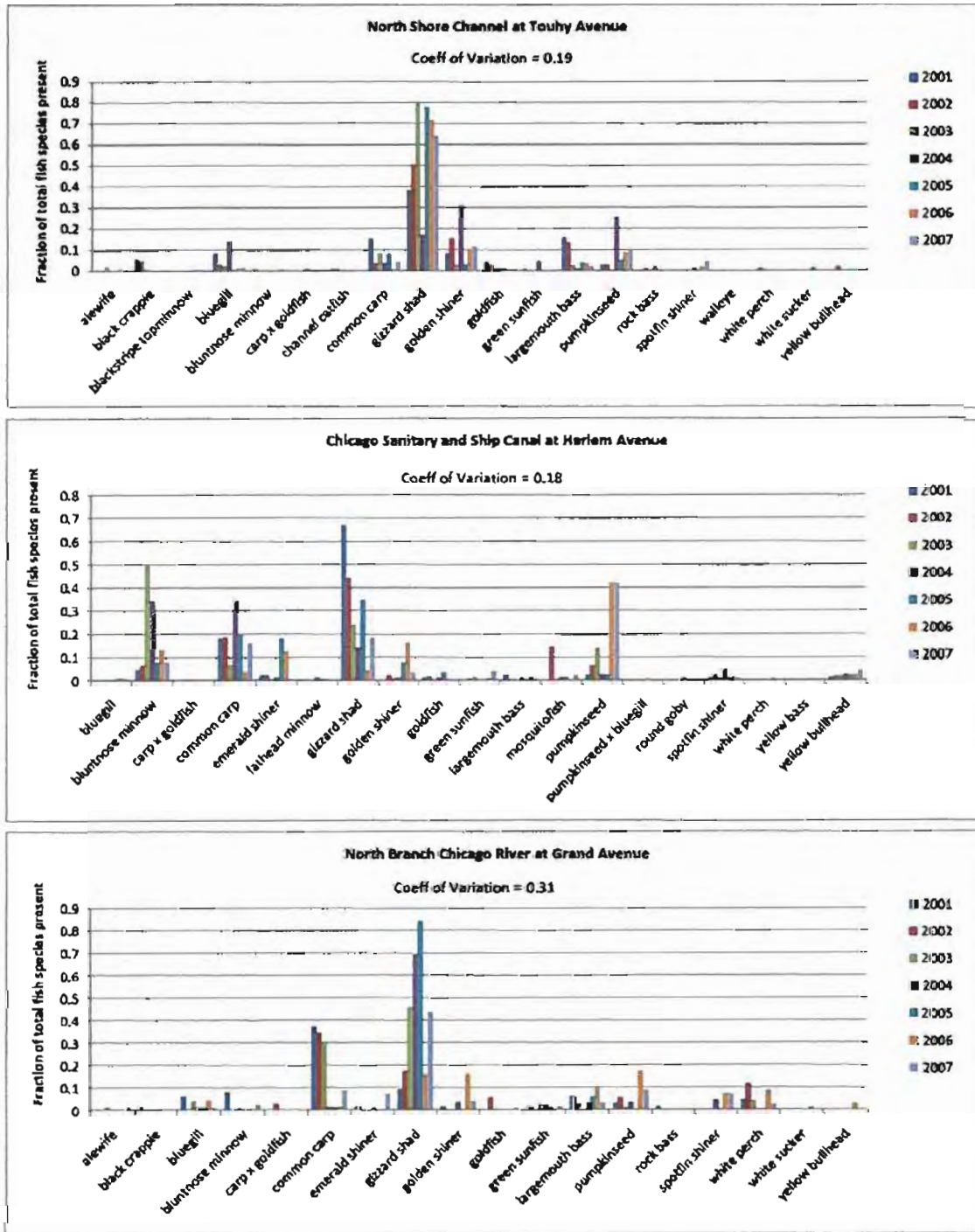


Figure 2-7. 2001 to 2007 Annual Station Fish Survey Results

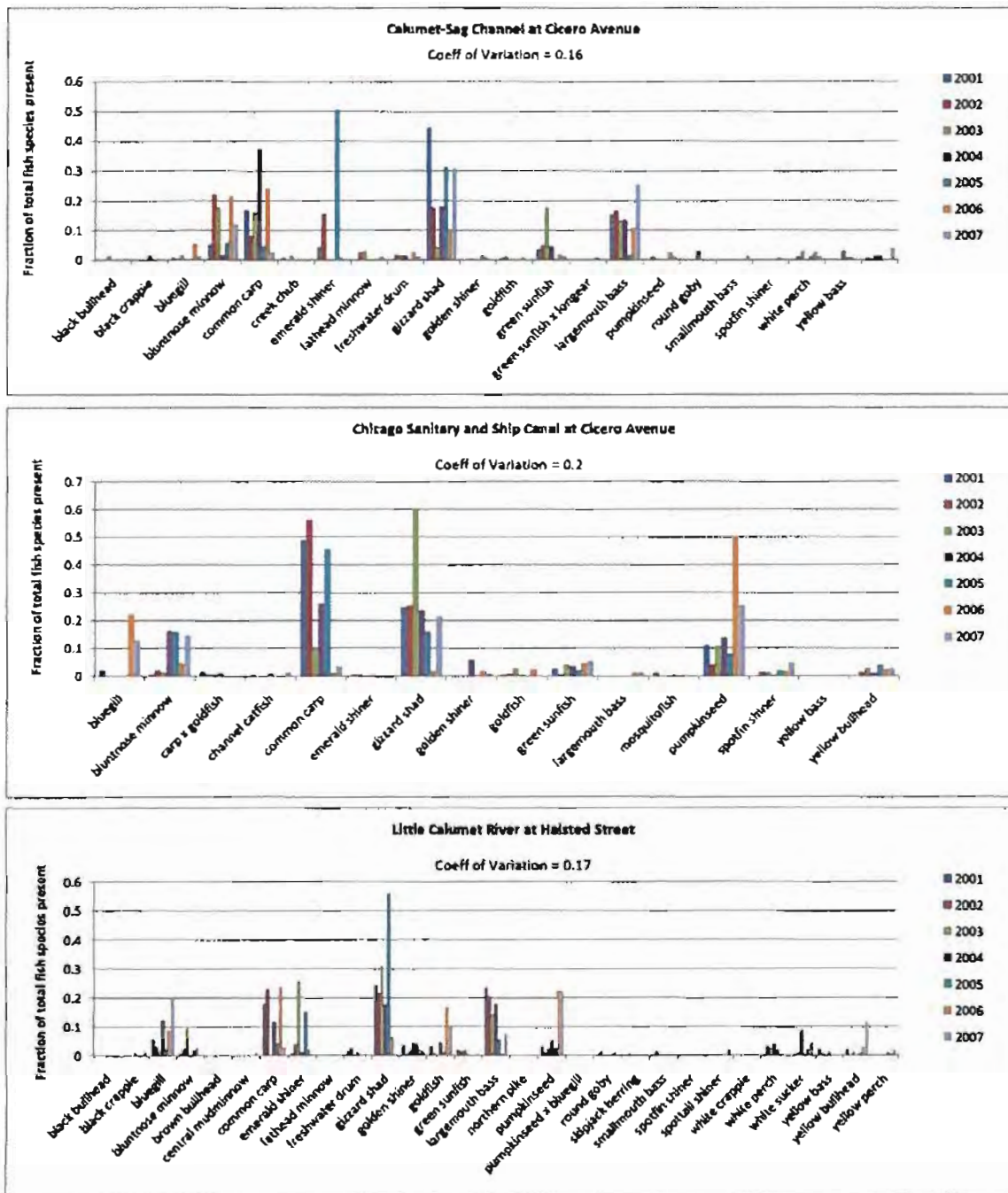


Figure 2-7. 2001 to 2007 Annual Station Fish Survey Results - Continued

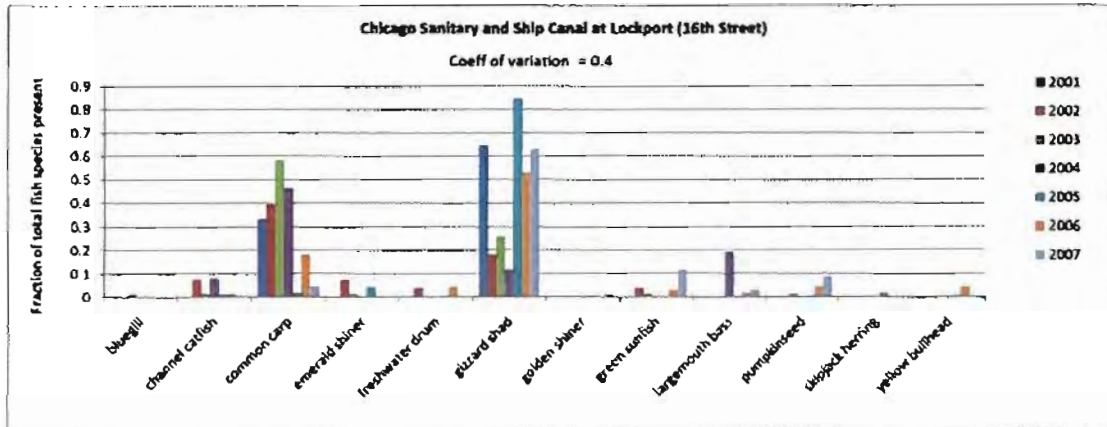


Figure 2-6. 2001 to 2007 Annual Station Fish Survey Results - Continued

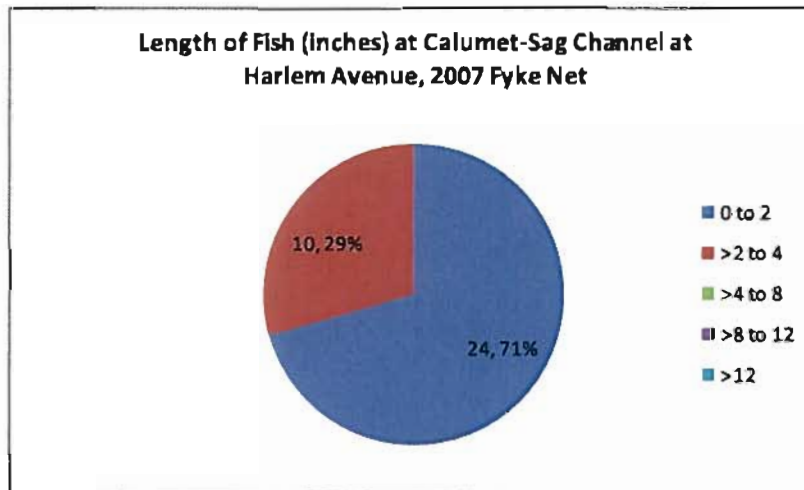
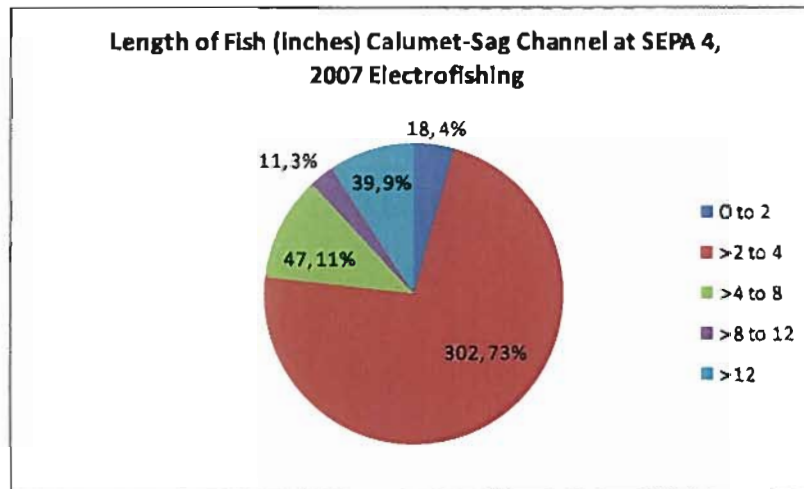
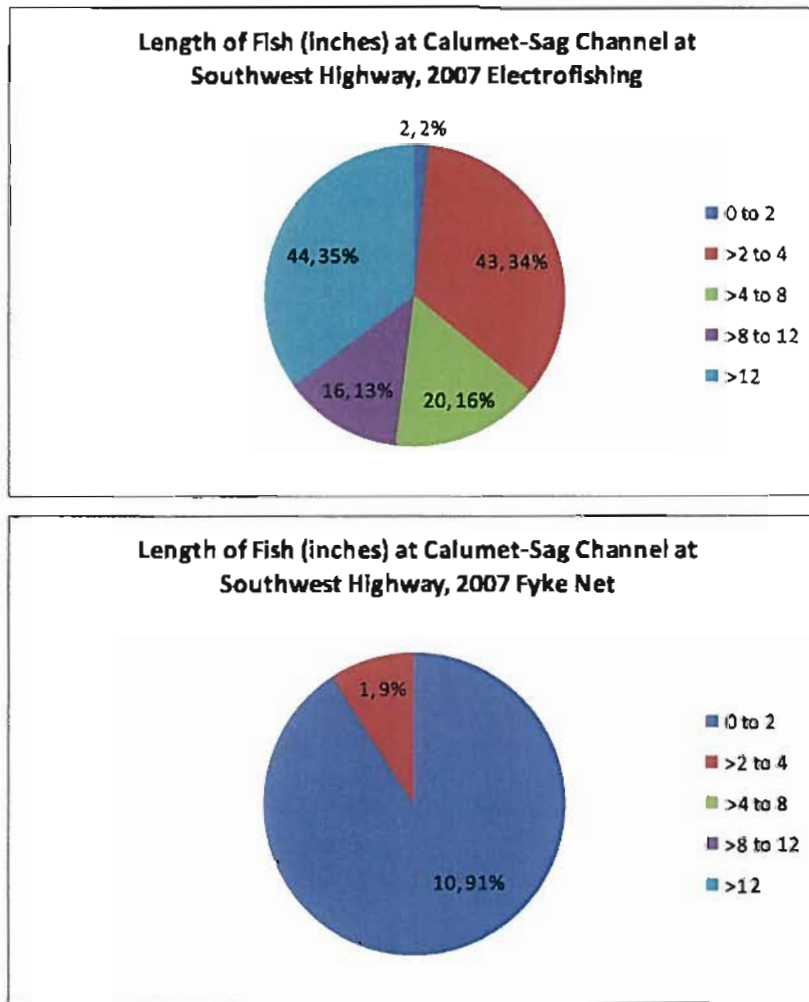


Figure 2-8. Results of Electrofishing and Fyke Net Samples by Length Interval, from 2007 Samples near Harlem Avenue on the Cal-Sag Channel.





**Figure 2-9. Results of Electrofishing and Fyke Net Samples by Length Interval ,  
from 2007 Samples near Southwest Highway on the Cal-Sag Channel.**

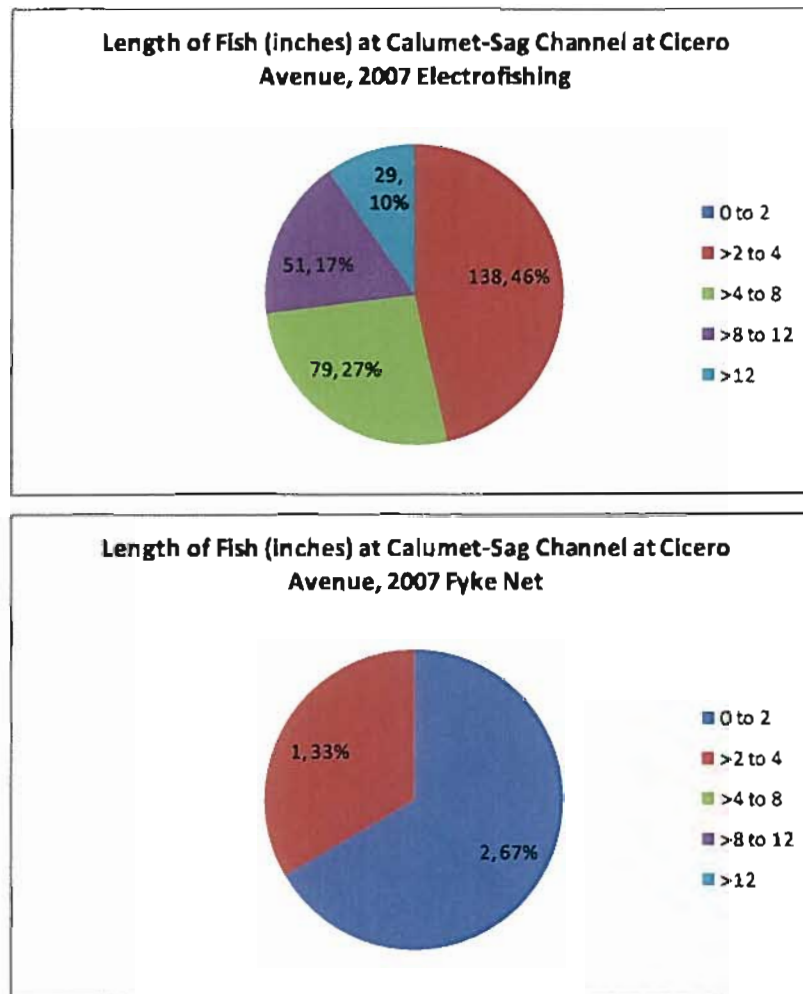


Figure 2-10. Results of Electrofishing and Fyke Net Samples by Length Interval ,  
from 2007 Samples near Cicero Avenue on the Cal-Sag Channel.



### 3. SELECTION OF FISH METRICS

Fish metric selection and calculation is a common form of fish data analysis (Flotemersch et al. 2006). The general approach for screening fish metrics to determine which will be most useful and appropriate for this study follows methods applied in development of fish IBIs, as documented in peer-reviewed scientific literature. As stated in the preceding section, the objective of this study is not to develop a new IBI for the CAWS, but the process of metric development involves review, analysis, and reduction of fish metrics, so the methods used in the literature to develop IBIs provides a sound basis for screening of metrics appropriate for the CAWS.

#### 3.1 COMPILATION OF FISH METRICS

Roset et al. (2007) suggests that starting with a large list of relevant candidate metrics increases the rigor of the system-specific metric selection process, by removing a level of *a priori* bias retained from previous studies. Lyons et al. (2001) provides a list of 26 fish metrics that were used as the starting point for the Wisconsin large warm water river IBI. The Lyons study is particularly relevant because it was developed in the Midwest for a range of larger river types, it is frequently cited, and Lyons' methodology is well-documented. Starting with Lyons' list of 26 fish metrics, LimnoTech then reviewed other relevant and significant IBI documents to identify other potentially applicable metrics:

- The Illinois IBI (IDNR 2000) was consulted as it currently provides the reference that the Illinois Environmental Protection Agency (IEPA) uses to determine attainment with aquatic life uses and may offer applicable metrics for the unique conditions within the CAWS (IEPA 2005). From this reference, ten additional metrics were added.
- The Ohio Boatable IBI (OEPA 1988) was consulted because it is frequently cited, still used after 20 years, one of the few fish IBI developed specifically for non-wadeable waters in the Midwest, and may offer applicable metrics for the unique conditions within the CAWS. Three additional metrics were included from the Ohio IBI.
- Karr's original work (Karr 1981) on fish IBIs was consulted because it was the seminal work on fish IBIs and most subsequent fish IBI work has been derived from it. No additional metrics were identified from this reference because they are included, as appropriate, in the above IBIs.

The metrics from the Illinois and Ohio IBIs increased the total number of metrics under consideration to 40. In addition to these previously used metrics, review of fish data from the CAWS and knowledge of the system suggested that some additional metrics would be worthy of consideration, including the following:

- *Percent intolerant species by number and by weight* – These metrics were added to provide additional quantification of the prevalence of pollution-sensitive individuals. This may provide information beyond the number of intolerant species.
- *Percent moderately tolerant species by number and by weight* – Previous studies have grouped species into tolerant or intolerant categories, however modifications to water quality standards recently proposed by the Illinois EPA have used the term “intermediately tolerant”, so the inclusion of metrics that reflect species that are moderately tolerant to water quality impacts may be useful.
- *Number of tolerant species* – This metric was included to provide a metric of direct comparison with the number of intolerant and moderately tolerant species.
- *Number of sunfish species, excluding largemouth bass* – This metric was added because sunfish metrics used in other IBIs either included all sunfish or excluded both smallmouth and largemouth bass. Because smallmouth bass are a cool water species and are less tolerant of anthropogenic impacts, it was desirable to include them, while excluding largemouth bass because of their wide distribution across the CAWS.

With the addition of these ‘custom’ metrics, the list of potential fish metrics for consideration in this Study totaled 46. Review of additional scientific literature did not identify any more applicable metrics for inclusion, suggesting that the starting metric list will provide the rigor suggested by Roset et al. (2007). The 46 fish metrics and their sources are listed in Table 3.1.

Table 3-1. Initial List of Fish Metrics.

Fish Metric	Metric Name	Source
%DELT_(n)	% Diseased or with eroded fins, lesions, or tumors	Lyons et al. (2001)
CPUE	catch per unit effort	"
WPUE	weight per unit effort	"
%LRIV_(n)	% large river species by count	"
%LRIV_(wt)	% large river species by weight	"
%RIV_(n)	% riverine species by count	"
%RIV_(wt)	% riverine species by weight	"
%RNDSCK_(n)	% round sucker species by count	"
%RNDSCK_(wt)	% round sucker species by weight	"
%TOL_(n)	% tolerant species by count	"
%TOL_(wt)	% tolerant species by weight	"
INT	number of intolerant species	"
RIV	number of riverine species	"
%LTHPL_(n)	% lithophilic spawners by count	"
%LTHPL_(wt)	% lithophilic spawners by weight	"
NAT	number of native species	"
SCKR	number of sucker species	"
SR	total number of species	"
SUN1	number of sunfish species, excluding smallmouth and largemouth bass	"
SUN2	number of sunfish species, including smallmouth and largemouth bass	"
%INSCT_(n)	% insectivores by count	"
%INSCT_(wt)	% insectivores by weight	"
%OMV_(n)	% omnivores by count	"
%OMV_(wt)	% omnivores by weight	"
%TC_(n)	% top carnivores by count	"
%TC_(wt)	% top carnivores by weight	"
PRTOL	proportion of Illinois tolerant species	IDNR, 2000
LITOT	IL ratio of non tolerant coarse-mineral-substrate spawners	"
INTOL	number of IL native intolerant species	"
NFSH	number of IL native species	"
NMIN	number of IL native minnow species	"
NSUC	number of IL native sucker species	"
NSUN	number of IL native sunfish species	"
GEN	IL ratio of generalist feeders	"
NBINV	IL native benthic invertivore species	"
SBI	IL ratio of specialist benthic invertivore species	"
TNI	total number of individuals	OEPA, 1988
OH_B_Sun	number of OH native sunfish species	"
%OH_B_OMN(n)	% OH omnivores, excluding channel catfish	"
%INT_(n)	% intolerant species by count	New for this Study
%INT_(wt)	% intolerant species by weight	"
%MOD_(n)	% moderately intolerant species by count	"
%MOD_(wt)	% moderately intolerant species by weight	"
MOD	number of moderately tolerant species	"
TOL	number of tolerant species	"
SUN3	number of sunfish species, excluding largemouth bass	"

### 3.2 SPECIFICATION OF TOLERANCE VALUES

Several of the metrics identified for screening are intended to be relative indicators of species tolerance to pollution and other human impacts. Therefore these metrics require that species be classified according to their pollution tolerance. This is significant because proposed water quality standards for the CAWS are defined in terms of maintaining aquatic-life populations of fish species that are tolerant, intermediately tolerant, and/or intolerant. It should be noted that the proposed water quality standards do not assign fish species to these tolerance categories, nor do they refer to sources from which to derive tolerance assignments.

The classification of fishes into tolerance categories has typically been based on best professional judgment (BPJ) assignments of species based on general responses to environmental degradation (Meador and Carlisle 2007). Meador and Carlisle (2007) cited that the relative success of BPJ classifications of tolerance in the Midwest may be a result of the perceived homogeneity of regional conditions and that the assignments may have limited geographic application. Further, tolerance assignments rarely discriminate among pollutant stressors. Meador and Carlisle (2007) found that stressors such as suspended sediment, conductivity, chloride and total phosphorus provided a better measure of pollution tolerance assignment than the typically considered stressors of temperature, dissolved oxygen and pH. For example, white sucker (*Catostomus commersoni*) and fathead minnow (*Pimephales promelas*) are generally categorized as tolerant to pollutants by Illinois DNR (IDNR 2000) and Meador and Carlisle (2007), despite their intolerance to low dissolved oxygen and high temperatures (Meador and Carlisle 2007). Unfortunately, the detailed stressor assignments by Meador and Carlisle (2007) have not been developed for the Midwest region and do not consider many of the CAWS species, so their method will not be used here, but warrants future consideration.

The approach for assigning CAWS species to pollution tolerance categories of tolerant, intolerant or moderately tolerant, attempted to rely on locally derived sources, although no single source covered all species found within the CAWS. The approach started with tolerance assignments established at the state level (IDNR 2008), then for the Midwest (Lyons et al. 2001), at the national level (Meador and Carlisle 2008) and then for specific references where a species was not included in the previous documents.

The State of Illinois has developed a manual for calculating fish IBIs that is in draft form with continued updates (IDNR 2000). The manual includes pollution tolerance assignments for a range of species. The IDNR (2000) assignments only include tolerant or intolerant for those with any assignment and most species in the state list have no assignment (that is, they are given a “—”). The classifications were derived from regional fish manuals including Smith (1979), Becker (1983), Karr et al. (1986), Jenkins and Burkhead (1994), Bertrand et al. 1996, OEPA (1988) and BPJ, where information was not available (IDNR 2008). These classifications were retained as a primary reference sources.

The next level of tolerance assignment was derived from Lyons et al. (2001). The Lyons paper provided additional assignments to some species not assigned by IDNR

(2008) but also restricted species assignments to tolerant or intolerant categories only, with the remaining species assigned as “other”. The tolerance assignments of Lyons stems from his earlier paper (Lyons 1992) where three qualitative criteria are used:

- 1) a known high degree of sensitivity to the types of environmental degradation as described by Becker (1983) and other regional fish publications;
- 2) areas of observed regions of decline in Wisconsin where environmental problems are known; and
- 3) designations used in other IBIs.

Meador and Carlisle (2007) from the U.S. Geological Survey (USGS) conducted an extensive analysis and assignment of numerous species into tolerant, moderately tolerant, and intolerant categories based on a recently published, quantified evaluation against physiochemical variables. The data set used for this effort is from the USGS national program and collected data from the USGS National Water Quality Assessment Program. These assignments were applied after the Lyons assignments. This effort resulted in a database of tolerance assignments for most remaining fish species, except for some remaining exotics. Finally, for those species not given tolerance assignments by the aforementioned efforts, species-specific papers were consulted and referenced for final pollution tolerance assignments. The tolerance values assigned for each species are included in Attachment B.

**This page is blank to facilitate double sided printing.**

## **4. SCREENING OF FISH METRICS**

The procedures and rationale for screening of fish metrics are described below.

### **4.1 SCREENING OBJECTIVES**

The process of screening the fish metrics had two primary objectives, as described below:

1. First, it was necessary to reduce the list of fish metrics to a more manageable number. Because the data corresponding to these metrics will be used for comparison to water quality and habitat data, too large a number of fish metrics would be too cumbersome. Metrics used to assess fishes vary based on the physical and biotic nature of the system (Flotemersch et al. 2006). Most fish IBIs reviewed for this study used a final set of ten to sixteen metrics (Karr 1981; OEPA 1988; Hughes et al., 1998; IDNR 2000; Lyons et al., 2001), so the goal was to reduce the list to within this range.
2. Second, the current scientific literature suggests that it is important to retain at least one metric from each major category of ecological function: species richness and composition, indicator species, trophic function, reproductive function, and individual abundance and condition (Simon and Lyons 1995; Lyons et al. 2001; Roset et al. 2007). Each category reflects a different aspect of fish assemblages that responds uniquely to aquatic ecosystem stressors (Hughes and Oberdorff 1999).

With these objectives in mind, the initial list of fish metrics was screened using the process described in the following sections.

### **4.2 METRICS LACKING DATA**

The initial step in the screening process was to identify metrics for which there were no data available. This was essential, because the metrics will eventually be used for statistical or other quantitative comparisons to other data types (i.e., water quality and habitat) and the lack of data would preclude such quantitative comparisons.

Review of the CAWS fish data from 2001 to 2007 revealed two metrics for which no data exist in the CAWS: the percentage of round sucker taxa (genera *Cycleptus*, *Hypentelium*, *Minytrema*, and *Moxostoma*) by weight and by number (%RNDSCCK\_(n) and %RNDSCCK\_(wt)). Based on this observation, these metrics were eliminated from further consideration. This initial screening reduced the list of fish metrics from 46 to 44.

### **4.3 METRIC RANGE**

Review of the scientific literature for fish IBI development shows that a typical method of screening fish metrics is to examine those metrics that reflect the number of species identified in a particular category or type and to screen out those that represent relatively

few species (McCormick et al. 2001; Emery et al. 2003). This so-called “range test” is used to eliminate metrics for which between 0 and 2 species were identified.

The “range test” was applied to the CAWS fish data and four metrics were found for which only one or two species were identified between 2001 and 2007. These four metrics were: the number of Illinois native benthic invertivore species (NBINV), the number of Illinois native sucker species (NSUC), the number of sucker species (SCKR), and the Illinois ratio of specialist benthic invertivore species (SBI). On the basis of this observation, these four metrics were eliminated from further consideration, reducing the number of potential metrics to 40.

#### **4.4 METRIC REDUNDANCY**

A very common method of screening metrics is to analyze the metrics for redundancy with each other. This method of screening is commonly used in index development (Hughes et al. 1998; Lyons et al. 2001, Emery et al. 2003, Wilhelm et al. 2005). In this analysis, Pearson’s correlation was calculated for pairs of metrics and the resulting correlation values were used to screen out statistically redundant metrics. This process is described in more detail below.

Before calculating the Pearson correlation coefficients, the metrics were evaluated for normality and several metrics were found to have skewed distributions. Some were right skewed, others were left skewed. The left skewed metrics were log transformed, resulting in near-normal distributions and include the following metrics: WPUE, TNI, TOL\_TNI, CPUE, %TC\_(wt), %LTHPL\_(wt), %MOD\_(n), and %MOD\_(wt). For the right skewed metrics (mostly data representing proportions) the arcsine-square-root-transform was evaluated, but because the distribution shapes did not improve these metrics were left untransformed.

Pearson correlation coefficients were calculated between the individual metrics in order to identify metrics that are highly correlated. Correlated metrics indicate some degree of redundancy, i.e. they respond similarly to characteristics of the CAWS system and can be used to derive similar conclusions. Threshold correlation strength had to be chosen to identify the metrics with “strong” correlation, as reported in the literature. In the literature reviewed, this threshold correlation value was usually between 0.6 and 0.75 (Lyons et al. 2001; McCormick et al. 2001; Emery et al. 2003; Whittier et al. 2007). For this analysis a value of 0.6 was used, which is what Lyons used for his large warm water river IBI (Lyons et al. 2001). Thus, pairs of metrics with a correlation coefficient above the threshold were defined as redundant and only one metric of the pair was retained for subsequent analyses. The matrix of Pearson’s correlation coefficients is presented in Attachment C.

Because many metrics were highly correlated with multiple metrics, some judgment was necessary in using this screening method to insure representation from each of the five ecological function categories. For example, the original list of 46 metrics only contained



three reproductive function metrics and five abundance and condition metrics, therefore these metrics were, in some cases, preferentially retained.

This screening step was successful in reducing the number of metrics from 40 to 16. The list of metrics remaining after screening for redundancy is presented in Table 4-1.

**Table 4-1. Fish Metrics Remaining after Screening for Redundancy.**

Fish Metric	Metric Name	Ecological Function Category
%DELT_(n)	% Diseased or with eroded fins, lesions, or tumors	ACM
CPUE	catch per unit effort	ACM
%LTHPL_(n)	% lithophilic spawners by count	RFM
%LTHPL_(wt)	% lithophilic spawners by weight	RFM
%INSCT_(n)	% insectivores by count	TFM
%INSCT_(wt)	% insectivores by weight	TFM
%TC_(n)	% top carnivores by count	TFM
%TC_(wt)	% top carnivores by weight	TFM
PRTOL	proportion of Illinois tolerant species	ISM
LITOT	IL ratio of non tolerant coarse-mineral-substrate spawners	RFM
INTOL	number of IL native intolerant species	ISM
NMIN	number of IL native minnow species	SRC
NSUN	number of IL native sunfish species	SRC
GEN	IL ratio of generalist feeders	TFM
%INT_(n)	% intolerant species by count	ISM
%MOD_(wt)	% moderately intolerant species by weight	ISM

#### 4.5 METRIC VARIABILITY

After applying the methods described above, the number of retained metrics (16) still exceeded the target number of metrics, so the retained metrics were inspected to determine whether a rational scientific basis could be identified for elimination of any of them.

It was noted that the set of metrics listed in Table 4-1 contained three pairs of metrics that represented similar fish attributes for both count and weight:

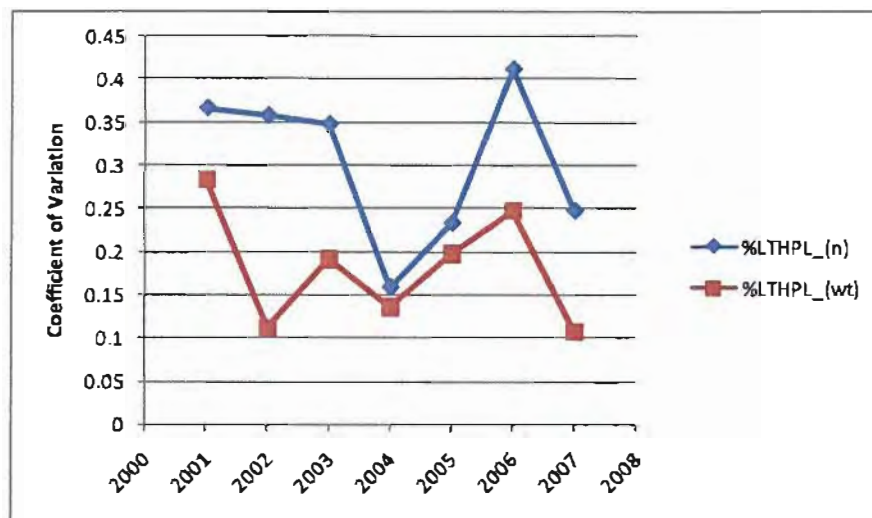
- % lithophilic spawners by count (%LTHPL\_(n)) and weight (%LTHPL\_(wt))
- % insectivores by count (%INSCT\_(n)) and weight (%INSCT\_(wt))
- % top carnivores count (%TC\_(n)) and weight (%TC\_(wt))

In addition, two metrics remained that represented intolerant species: %INT\_(n) and INTOL. Because each of these four pairs of metrics measure the same attributes of fish assemblages, it seemed appropriate to select one metric from each pair to carry forward. To determine which metric in each pair to retain, the variability of the metrics within the data set was examined. The rationale for using metric variability as a screening measure was that preference should be given to metrics that exhibited greater variation within the

system, since those metrics will be more likely to help identify relationships to other system attributes such as water quality and physical habitat.

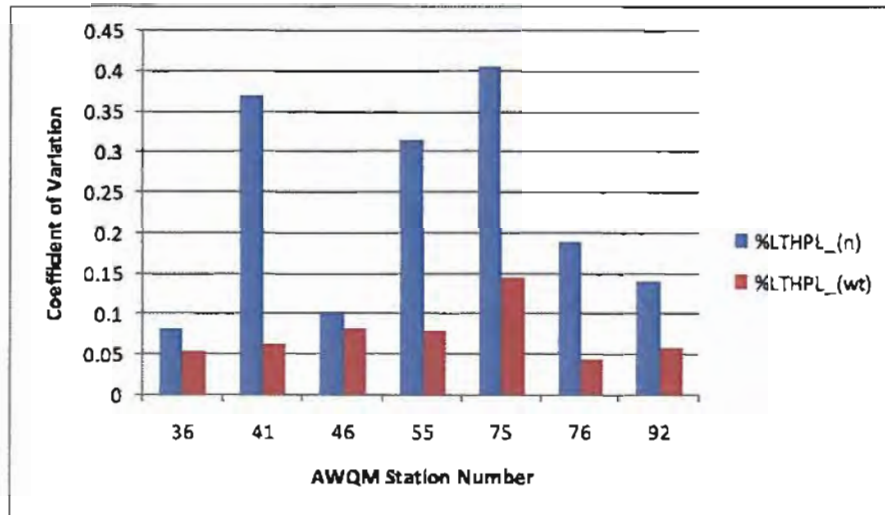
Calculated values for each of the paired metrics were extracted from the CAWS fish database and the coefficient of variation (CV) for each metric was calculated using all data from each year from 2001 through 2007 to give a measure of data variability in each year for each metric. The CV for each metric was also calculated at each of seven annual sampling stations for all years to determine variability across the system. The results are discussed below.

The system-wide CVs for %LTHPL\_(wt) and %LTHPL\_(n) are depicted graphically in Figure 4-1.



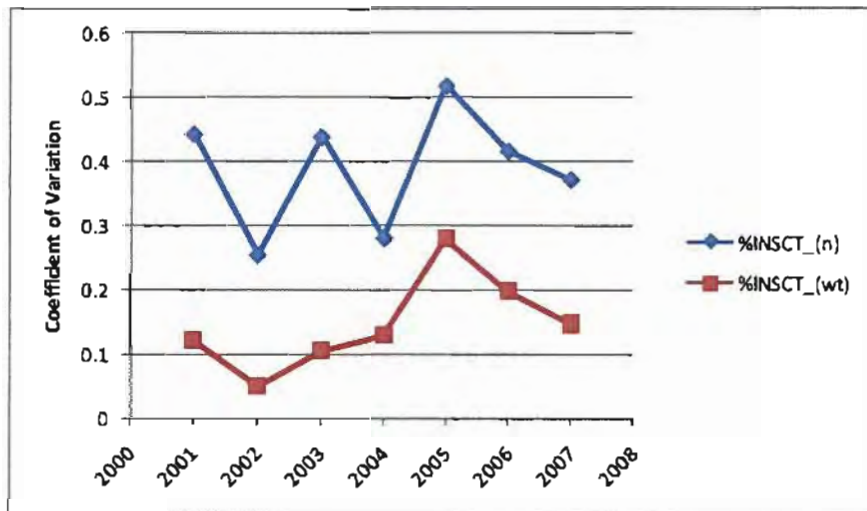
**Figure 4-1. Coefficient of Variation for %LTHPL\_(wt) and %LTHPL\_(n), for 2001 through 2007 Data.**

Although the CVs for both %LTHPL\_(wt) and %LTHPL\_(n) are both very low (less than 0.5 in every year), the calculated value for %LTHPL\_(n) is consistently higher, in many cases double that of %LTHPL\_(wt). The CVs for %LTHPL\_(n) also appear to exhibit more variability over time than for %LTHPL\_(wt), which is also evident from the CVs calculated for the annual sampling stations depicted in Figure 4-2. Based on these observations, %LTHPL\_(n) was retained and %LTHPL\_(wt) was eliminated.

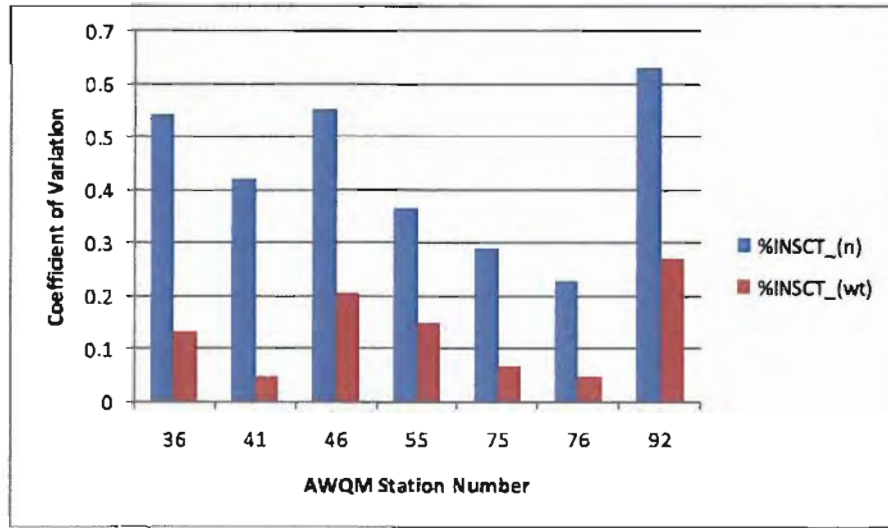


**Figure 4-2. Coefficient of Variation for %LTHPL\_(wt) and %LTHPL\_(n) at Annual Sampling Stations.**

The same comparison was made for %INSCT\_(n) and %INSCT\_(wt). In this case, the CV for %INSCT\_(n) is consistently higher than for %INSCT\_(wt), both on a system-wide basis across multiple years (Figure 4-3) as well as when compared between annual sampling stations (Figure 4-4). On the basis of these comparisons, %INSCT\_(n) was retained and %INSCT\_(wt) was eliminated.



**Figure 4-3. Coefficient of Variation for %INSCT\_(wt) and %INSCT\_(n), for 2001 through 2007 Data.**



**Figure 4-4. Coefficient of Variation for %INSCT\_(wt) and %INSCT\_(n) at Annual Sampling Stations.**

Similarly, the CVs for %TC\_(n) and %TC\_(wt) were compared. In the case of this metric pair, most of the CVs for %TC\_(wt) were above 1.0, while all the CVs for %TC\_(n) were below 1.0, suggesting that %TC\_(wt) has significantly higher variability (Figure 4-5). While some sampling stations exhibited similar CVs for both %TC\_(wt) and %TC\_(n) (Figure 4-6), three stations had significantly higher CVs for %TC\_(wt). Based on these observations, %TC\_(wt) was retained and %TC\_(n) was eliminated.

Finally, the CVs for %INT\_(n) and INTOL were compared both on a system-wide basis for each sampling year and for each annual sampling station across all years. The comparison of system-wide variability through time (Figure 4-7) clearly indicates that %INT\_(n) has higher variability than INTOL, even though the inter-station comparison (Figure 4-8) shows similarity between the two metrics in terms of variability. On the basis of these observations, %INT\_(n) was retained and INTOL was eliminated.

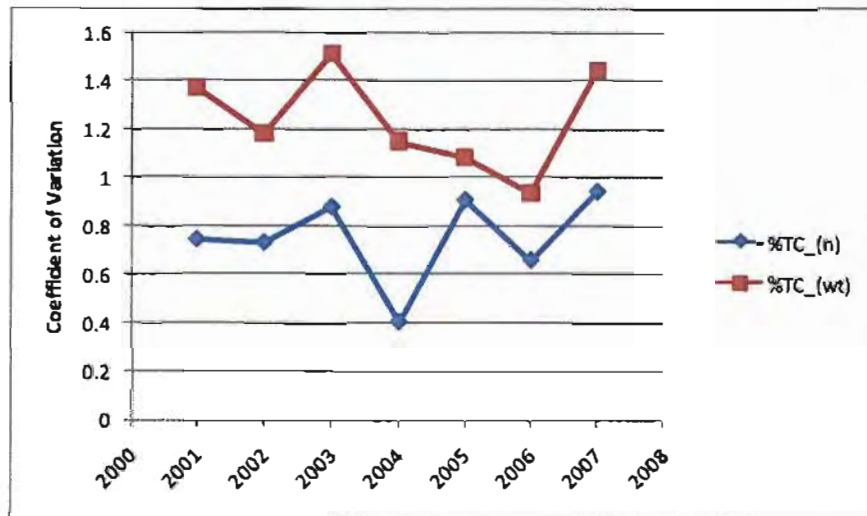


Figure 4-5. Coefficient of Variation for %TC\_(wt) and % TC\_(n), for 2001 through 2007 Data.

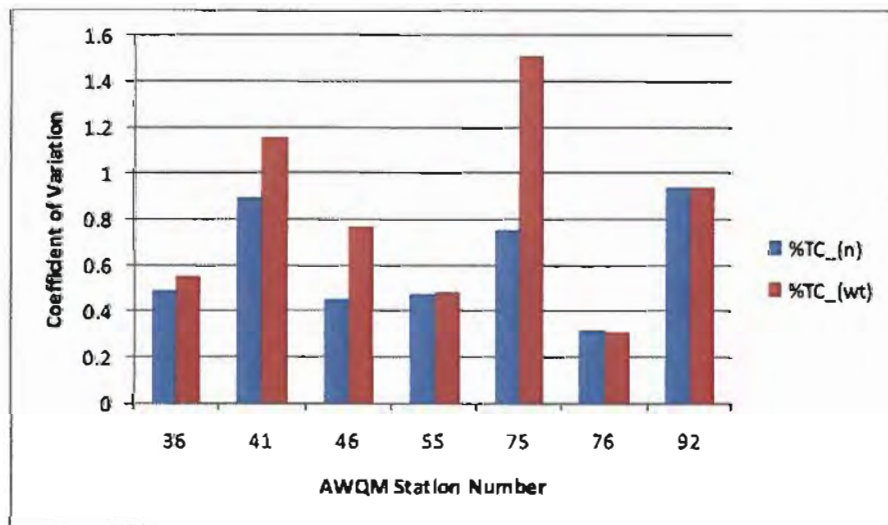


Figure 4-6. Coefficient of Variation for %TC\_(wt) and % TC\_(n) at Annual Sampling Stations.

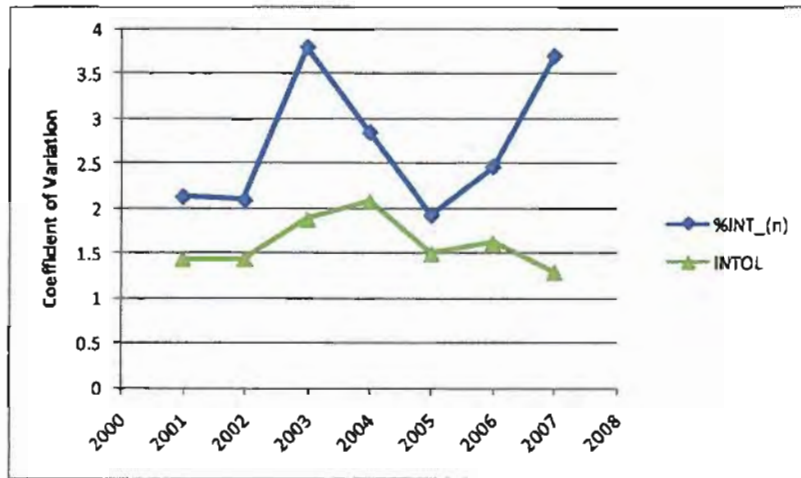


Figure 4-7. Coefficient of Variation for %INT\_(n) and INTOL, for 2001 through 2007 Data.

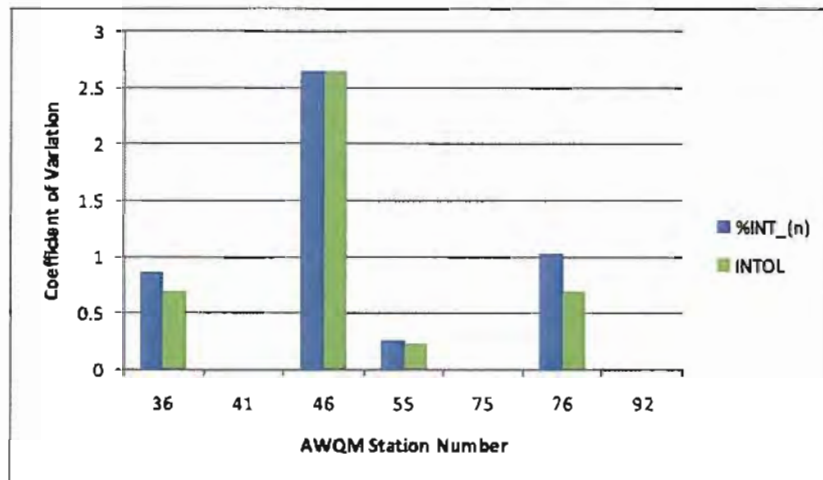


Figure 4-8. Coefficient of Variation for %INT\_(n) and INTOL at Annual Sampling Stations.

In summary, based on review of metric variability as quantified by each metric's coefficient of variation, the following metric selections were made:

- %LTHPL\_(n) was retained over %LTHPL\_(wt);
- %INSCT\_(n) was retained over %INSCT\_(wt);
- %TC\_(wt) was retained over %TC\_(n); and
- %INT\_(n) was retained over INTOL.

These selections reduced the list of metrics to 12, which are summarized in the following section.



## 5. FINAL RECOMMENDED LIST OF METRICS

After completion of the screening process described in the preceding section, twelve metrics were retained for use in the CAWS (Table 5-1). The retained metrics are representative of each of the five ecological function categories as recommended by Simon and Lyons (1995), Lyons et al. (2001), Roset et al. (2007): species richness and composition (SRC), indicator species (ISM), trophic function (TFM), reproductive function (RFM), and individual abundance and condition (ACM). These are further described below,

SRC category includes two native species metrics. Species richness and composition are a measure of species diversity and Hughes and Oberdorff (1999) suggest using native species metrics for assessing physical or water quality stressors where non-natives are abundant, as found in the CAWS. Both metrics are also used by the State of Illinois and should be appropriate measures for species richness assessments within the CAWS.

ISM includes three proportional metrics of tolerant, moderately tolerant and intolerant measures. Proportional measures for species have been recommended by others as well (Karr et al. 1986; Lyons et al. 1995). The current numbers of intolerant species across the CAWS is generally low and it is generally expected that the proportion of intolerant species is responding to physical and water quality stressors unique to the CAWS. However, it is expected that these species would respond positively to stressor reductions and may provide an appropriate metric for the CAWS. Both tolerant and moderately tolerant species are wide-spread across the CAWS and it is assumed that the tolerant metrics would respond negatively to physical and water quality improvements while moderately tolerant species proportions increase with the reduction of stressors. All three proportional measures are applicable measures across the CAWS.

TFM includes a range of feeding metrics for the CAWS that include top carnivores, generalists and insect feeders. It is generally expected that top carnivores and insectivores would respond negatively to physical and water quality stressors, while generalists would respond positively to these stressors (Flotemersch et al. 2006). All three metrics are applicable across the CAWS, are appropriate measures of trophic function and are supported by the original work of Karr (1981) and subsequent authors (Hughes and Oberdorff 1999).

RFM includes a proportion of all lithophilic species as well as intolerant lithophilic species native to Illinois. It is generally expected that lithophilic species would respond negatively to both physical and water quality stressors (Flotemersch et al. 2006). Although it is expected that lithophilic habitat is limited across the CAWS, these metrics are included because existing habitat conditions as well as future improvements within portions of the CAWS should result in a positive response by these metrics. The metrics are used within the Illinois IBI as

well as others (Emery et al. 1999; Flotemersch et al. 2006) and are appropriate for the CAWS.

ACM includes a metric for the condition of the sampled fishes as well as the efficiency of the collection methods. It is generally expected that the observed number of physical anomalies of collected fishes changes in response to a range of water quality stressors. Hughes and Oberdorff (1999) suggest including this metric where the possibility for changes in the incidence of disease and deformity exist. Hughes and Oberdorff (1999) describe sample abundance as a surrogate for system productivity but caution that nutrient and thermal enrichment may affect this metric response. Typically, it is expected that the efficiency of collected fishes decrease in response to both water quality and habitat stressors (Flotemersch et al. 2006) but the uniqueness of the CAWS conditions may warrant special consideration of the use of this metric in subsequent analysis. Both measures are commonly used measures for ACM and are appropriate for the CAWS.

In summary, the methods used for fish metric selection for the CAWS are appropriate, literature supported and robust methods. These methods have produced a final metric list that is appropriate and sensitive to responses of both physical habitat and water quality conditions within the CAWS and will be useful for further fish-habitat and fish-water quality analyses.

**Table 5-1. Final Recommended Fish Metrics for Use in the CAWS.**

Fish Metric	Metric Name	Ecological Function Category
%DELT_(n)	% Diseased or with eroded fins, lesions, or tumors	ACM
CPUE	catch per unit effort	ACM
%LTHPL_(n)	% lithophilic spawners by count	RFM
%INSCT_(n)	% insectivores by count	TFM
%TC_(wt)	% top carnivores by weight	TFM
PRTOL	proportion of Illinois tolerant species	ISM
LITOT	IL ratio of non tolerant coarse-mineral-substrate spawners	RFM
NMIN	number of IL native minnow species	SRC
NSUN	number of IL native sunfish species	SRC
GEN	IL ratio of generalist feeders	TFM
%INT_(n)	% intolerant species by count	ISM
%MOD_(wt)	% moderately intolerant species by weight	ISM



## 6. REFERENCES

- Becker, G. C., ed., 1983. *Fishes of Wisconsin*, The University of Wisconsin Press. Madison, Wisconsin.
- Bertrand, W.A., R.L. Hite and D.M. Day. 1996. Biological Stream Characterization: Biological Assessment of Illinois Stream Quality through 1993. IEPA. IEPA/BOW/96-058. Springfield, Illinois.
- Bradley, D. L., 2008. "Field Work Correspondence With Anglers During 2008 Field Work Within the Chicago Area Waterway System. Doug Bradley (LimnoTech, Inc.) June - September 2008."
- Emery, E. B., Simon, T. P., McCormick, F. H., Angermeier, P. L., Deshon, J. E., Yoder, C. O., Sanders, R. E., Pearson, W. D., Hickman, G. D., Reash, R. J., and Thomas, J. A., 2003. "Development of a Multimetric Index for Assessing the Biological Condition of the Ohio River," *Transactions of the American Fisheries Society*, Vol. 132, pp. 791-808.
- Flotemersch, J. E., Stribling, J. B., and Paul, M. J., 2006. "Concepts and Approaches for the Bioassessment of Non-Wadeable Streams and Rivers." Rep. No. EPA 600-R-06-127, US Environmental Protection Agency. Cincinnati, Ohio.
- Hubert, W. A., 1996. "Passive Capture Techniques," *Fisheries Techniques*. Brian R. Murphy, and David W. Willis, eds., American Fisheries Society, Bethesda, Maryland. pp. 157-181.
- Hughes, R. M., Kaufmann, P. R., Herlihy, A. T., Kincaid, T. M., Reynolds, L., and Larson, D. P., 1998. "A Process for Developing and Evaluating Indices of Fish Assemblage Integrity," *Canadian Journal of Fisheries and Aquatic Sciences*, Vol. 55, pp. 1618-1631.
- Hughes, R. M., and Oberdorff, T., 1999. "Applications of IBI Concepts and Metrics to Waters Outside the United States and Canada," *Assessing the Sustainability and Biological Integrity of Water Resources using Fish Communities*. T. P. Simon, ed., CRC Press, Boca Raton, Florida. pp. 79-93.
- IDNR, 2000. "Draft Manual for Calculating Index of Biotic Integrity Scores for Streams in Illinois." Illinois Department of Natural Resources.
- IDNR, 2008. "Personal Communication Between R. Smogor (IDNR) and D. Bradley (LimnoTech) Regarding Background of Illinois IBI Development."
- IEPA, 2005. "Interpreting Illinois Fish-IBI Scores, DRAFT." Illinois Environmental Protection Agency.

- Jenkins, R. E., and Burkhead, N. M., 1994. "Freshwater Fishes of Virginia." American Fisheries Society. Bethesda, Maryland.
- Karr, J. R., 1981. "Assessment of Biotic Integrity Using Fish Communities," *Fisheries*, Vol. 6, No.6., pp. 21-27.
- Karr, J. R., Fausch, K. D., Angermeier, P. L., Yant, P. R., and Schlosser, I. J., 1986. "Assessing Biological Integrity in Running Waters: A Method and Its Rationale." Illinois Natural History Survey Special Publication 5.
- Karr, J. R., 1991. "Biological Integrity: A Long-Neglected Aspect of Water Resource Management," *Ecological Applications*, Vol. 1, No.1., pp. 66-84.
- Lyons, J., 1992. "Using the Index of Biotic Integrity (IBI) to Measure Environmental Quality in Warmwater Streams of Wisconsin." Rep. No. NC-149, US Department of Agriculture. St. Paul, Minnesota.
- Lyons, J., Navarro-Perez, S., Cochran, P. A., Santana, E., and Guzman-Arroyo, M., 1995. "Index of Biotic Integrity Based on Fish Assemblages for the Conservation of Streams and Rivers in West-Central Mexico," *Conservation Biology*, Vol. 9, pp. 569-584.
- Lyons, J., Piette, R. R., and Niermeyer, K. W., 2001. "Development, Validation, and Application of a Fish-Based Index of Biotic Integrity for Wisconsin's Large Warmwater Rivers," *Transactions of the American Fisheries Society*, Vol. 130, pp. 1077-1094.
- McCormick, F. H., Hughes, R. M., Kaufmann, P. R., Herlihy, A. T., Peck, D. V., and Stoddard, J. L., 2001. "Development of an Index of Biotic Integrity for the Mid-Atlantic Highlands Region," *Transactions of the American Fisheries Society*, Vol. 130, pp. 857-877.
- Meador, M. R., and McIntyre, J. P., 2003. "Effects of Electrofishing Gear Type on Spatial and Temporal Variability in Fish Community Sampling," *Transactions of the American Fisheries Society*, Vol. 132, pp. 709-716.
- Meador, M. R., and Carlisle, D. M., 2007. "Quantifying Tolerance Indicator Values for Common Stream Fish Species of the United States," *Ecological Indicators*, Vol. 7, pp. 329-338.
- OEPA, 1988. "Biological Criteria for the Protection of Aquatic Life: Volume II: Users Manual for Biological Field Assessment of Ohio Surface Waters." Ohio Environmental Protection Agency. October 30, 1987 (Updated January 1, 1988).
- Paller, M. H., 1995. "Relationships Among Number of Fish Species Sampled, Reach Length Surveyed, and Sampling Effort in South Carolina Coastal Plain Streams," *North American Journal of Fisheries Management*, Vol. 15, pp. 110-120.

- Roset, N., Grenouillet, G., Goffaux, D., Pont, D., and Kestemont, P., 2007. "A Review of Existing Fish Assemblage Indicators and Methodologies," *Fisheries Management and Ecology*, Vol. 14, pp. 393-405.
- Simon, T. P., and Lyons, J., 1995. "Application of the Index of Biotic Integrity to Evaluate Water Resource Integrity in Freshwater Ecosystems," *Biological Assessment and Criteria: Tools for Water Resource Planning and Decision Making*. W. S. Davis, and T. P. Simon, eds., Lewis Publishers, pp. 245-262.
- Simon, T. P., and Sanders, R. E., 1999. "Applying an IBI Based on Great-River Fish Communities," *Assessing the Sustainability and Biological Integrity of Water Resources Using Fish Communities*. T.P.Simon, ed., CRC Press, Boca Raton, Florida. pp. 475-505.
- Smith, P. W., 1979. "The Fishes of Illinois." University of Illinois Press. Urbana, Illinois.
- Whittier, T. R., Hughes, R. M., Stoddard, J. L., Lomnický, G. A., Peck, D. V., and Herlihy, A. T., 2007. "A Structured Approach for Developing Indices of Biotic Integrity: Three Examples From Streams and Rivers in the Western USA," *Transactions of the American Fisheries Society*, Vol. 136, pp. 718-735.
- Wilhelm, J. G. O., Allan, J. D., Wessell, K. J., Merritt, R. W., and Cummins, K. W., 2005. "Habitat Assessment of Non-Wadeable Rivers in Michigan," *Environmental Management*, Vol. 36, No.4., pp. 592-609.

**This page is blank to facilitate double sided printing.**

**ATTACHMENT A:**

**CAWS FISH DATA STATIONS AND SAMPLING DATES**

**USED IN THIS ANALYSIS**

**This page is blank to facilitate double sided printing.**

EPA Station Description	Station ID	Station Description	2001	2002	2003	2004	2005	2006	2007	Total Count
North Shore Channel	35	North Shore Channel at Central Street	9/24/01				7/23/05			2
	38	North Shore Channel at Touhy Avenue	9/29/01	7/31/02	7/24/03	9/29/04	7/21/05	7/10/06	7/12/07	7
	101	North Shore Channel at Foster Avenue	9/27/01				9/9/05			2
North Branch Chicago River from its confluence with North Shore Channel to the south end of the North Avenue Turning Basin	102	North Shore Channel at Oakton Street	9/25/01				7/23/05			2
	37	North Branch Chicago River at Wilson Avenue	10/1/01				9/7/05			2
Little Calumet River from its confluence with Calumet River and Grand Calumet River to its confluence with Calumet-Sag Channel	73	North Branch Chicago River at Diversey Parkway	10/3/01				9/9/05			2
	56	Little Calumet River at Indiana Avenue			9/29/03				7/30/07	2
	76	Little Calumet River at Halsted Street	9/12/01	9/16/02	9/25/03	9/30/04	9/27/05	7/21/06	7/31/07	7
	902,SEPA2	Little Calumet River at SEPA 2					10/23/05	10/29/06		2
Calumet-Sag Channel	43	Calumet-Sag Channel at Route 83			7/20/03				9/14/07	2
	58	Calumet-Sag Channel at Ashland Avenue			9/5/03				9/1/07	2
	59	Calumet-Sag Channel at Cicero Avenue	9/14/01	9/17/02	7/31/03	8/31/04	9/29/05	7/24/06	9/29/07	7
	903,SEPA3	Calumet-Sag Channel at SEPA 3			10/6/03		10/23/05		10/51/07	3
	904,SEPA4	Calumet-Sag Channel at SEPA 4			10/3/03	10/18/04	10/16/05	10/30/06	10/29/07	5
	905,SEPA5	Calumet-Sag Channel at SEPA 5			10/1/03	10/18/04	10/16/05	10/17/06	10/29/07	5
		Calumet-Sag Channel at 104th Street							9/14/07	1
		Calumet-Sag Channel at Kedzie Avenue							9/13/07	1
	Calumet-Sag Channel at Southwest Highway							9/13/07	1	
North Branch Chicago River from the south end of the North Avenue Turning Basin to its confluence with South Branch Chicago River and Chicago River	46	North Branch Chicago River at Grand Avenue	10/2/01	8/1/02	7/23/03	8/27/04	7/18/05	7/11/06	7/11/07	7
Chicago River	74	Chicago River at Lake Shore Drive		8/2/02				7/29/06		2
	100	Chicago River at Wells Street		8/21/02				7/27/05		2
South Branch Chicago River and its South Fork	39	South Branch Chicago River at Madison Street		8/27/02				7/28/06		2
	40	Chicago Sanitary and Ship Canal at Damen Avenue		8/19/02				8/30/06		2
	99	Bubbly Creek at Archer Avenue		8/20/02				9/5/06		2
	106	South Branch Chicago River at Loomis Street		8/28/02				9/12/06		2
	98.2	Bubbly Creek at 35th St.			9/30/03	10/20/04	8/10/05			3
	99.1	Bubbly Creek at I-55			9/30/03	10/20/04	8/10/05			3
	99.3	Bubbly Creek at RAPS			9/30/03	10/20/04	8/10/05			3
Chicago Sanitary and Ship Canal	41	Chicago Sanitary and Ship Canal at Harlem Avenue	9/7/01	9/3/02	7/21/03	8/24/04	8/26/05	8/21/06	7/16/07	7
	42	Chicago Sanitary and Ship Canal at Route 83		8/28/02				8/31/06		2
	48	Chicago Sanitary and Ship Canal at Stephen Street		9/10/02				8/29/06		2
	75	Chicago Sanitary and Ship Canal at Cicero Avenue**	9/4/01	8/29/02	7/18/03	8/23/04	8/22/05	8/29/06	7/17/07	7
	92	Chicago Sanitary and Ship Canal at Lockport (16th Street)	9/4/01	9/11/02	7/29/03	8/30/04	9/15/05	7/25/06	7/10/07	7
	905.1,SEPA5_CS,SC	Chicago Sanitary and Ship Canal at SEPA 5			10/1/03	10/18/04	10/16/05	10/17/06	10/29/07	5
Total Stations 34			12	15	17	13	20	19	17	113

This page is blank to facilitate double-sided printing.



**ATTACHMENT B:**

**LIST OF FISH SPECIES IDENTIFIED IN THE CAWS (2001-2007)  
AND THEIR TOLERANCE ASSIGNMENTS**

**This page is blank to facilitate double sided printing.**

Scientific Name	Common Name	Tolerant	Intolerant	Moderate
<i>Alosa pseudoharengus</i>	alewife	X <sup>4</sup>		
<i>Alosa chrysochloris</i>	skipjack herring			X <sup>5</sup>
<i>Dorosoma cepedianum</i>	gizzard shad	X <sup>3</sup>		
<i>Oncorhynchus mykiss</i>	rainbow trout		X <sup>3</sup>	
<i>Umbra limi</i>	central mudminnow	X <sup>2</sup>		
<i>Esox lucius</i>	northern pike			X <sup>3</sup>
<i>Carassius auratus</i>	goldfish	X <sup>1</sup>		
<i>Cyprinus carpio</i>	common carp	X <sup>1</sup>		
<i>Notemigonus crysoleucos</i>	golden shiner	X <sup>1</sup>		
<i>Semotilus atromaculatus</i>	creek chub	X <sup>1</sup>		
<i>Cyprinella spiloptera</i>	spotfin shiner			X <sup>3</sup>
<i>Pimephales promelos</i>	fathead minnow	X <sup>1</sup>		
<i>Pimephales notatus</i>	bluntnose minnow	X <sup>1</sup>		
<i>Notropis atherinoides</i>	emerald shiner	X <sup>3</sup>		
<i>Notropis hudsonius</i>	spottail shiner		X <sup>2</sup>	
<i>Notropis stramineus</i>	sand shiner			X <sup>3</sup>
<i>Ictiobus niger</i>	black buffalo		X <sup>2</sup>	
<i>Catostomus commersoni</i>	white sucker	X <sup>1</sup>		
<i>Ictalurus punctatus</i>	channel catfish	X <sup>3</sup>		
<i>Amelurus natalis</i>	yellow bullhead	X <sup>1</sup>		
<i>Ameiurus melas</i>	black bullhead			X <sup>3</sup>
<i>Ameiurus nebulosus</i>	brown bullhead			X <sup>3</sup>
<i>Noturus gyrinus</i>	tadpole madtom			X <sup>3</sup>
<i>Fundulus notatus</i>	blackstripe topminnow	X <sup>3</sup>		
<i>Gambusia affinis</i>	mosquitofish	X <sup>3</sup>		
<i>Labidesthes sicculus</i>	brook silverside			X <sup>3</sup>
<i>Morone saxatilis</i>	striped bass			X <sup>6</sup>
<i>Morone chrysops</i>	white bass	X <sup>3</sup>		
<i>Morone mississippiensis</i>	yellow bass			X <sup>9</sup>
<i>Morone americana</i>	white perch	X <sup>7</sup>		
<i>Pomoxis nigromaculatus</i>	black crappie	X <sup>3</sup>		
<i>Pomoxis annularis</i>	white crappie	X <sup>3</sup>		
<i>Ambloplites rupestris</i>	rock bass		X <sup>2</sup>	
<i>Micropterus salmoides</i>	largemouth bass	X <sup>3</sup>		
<i>Micropterus dolomieu</i>	smallmouth bass		X <sup>1</sup>	
<i>Lepomis gulosus</i>	warmouth			X <sup>3</sup>
<i>Lepomis cyanellus</i>	green sunfish	X <sup>1</sup>		
<i>Lepomis macrochirus</i>	bluegill			X <sup>3</sup>
<i>Lepomis gibbosus</i>	pumpkinseed			X <sup>3</sup>
<i>Lepomis humilis</i>	orangespotted sunfish	X <sup>3</sup>		
<i>Stizostedion vitreum</i>	walleye			X <sup>3</sup>

Scientific Name	Common Name	Tolerant	Intolerant	Moderate
<i>Perca flavescens</i>	yellow perch			X <sup>3</sup>
<i>Aplodinotus grunniens</i>	freshwater drum	X <sup>3</sup>		
<i>Neogobius melanostomus</i>	round goby	X <sup>8</sup>		
<i>Cyprinus spp.</i>	carp x goldfish	X <sup>11</sup>		
<i>Oncorhynchus tshawytscha</i>	chinook salmon		X <sup>9</sup>	
<i>Oncorhynchus kisutch</i>	coho salmon		X <sup>9</sup>	
<i>Lepomis spp.</i>	green sunfish x bluegill	X <sup>11</sup>		
<i>Lepomis spp.</i>	green sunfish x longear	X <sup>11</sup>		
<i>Lepomis spp.</i>	green sunfish x pumpkinseed	X <sup>11</sup>		
<i>Oreochromis niloticus</i>	nile tilapia	X <sup>10</sup>		
<i>Lepomis spp.</i>	pumpkinseed x bluegill	X <sup>11</sup>		

**References**X<sup>1</sup> - IDNR 2000X<sup>2</sup> - Lyons et al. 2001X<sup>3</sup> - USGS 2008X<sup>4</sup> - FWS 1986X<sup>5</sup> - Barbour et al. 1999X<sup>6</sup> - EPA 2008X<sup>7</sup> - FWS 1983X<sup>8</sup> - Corkum et al. 2004X<sup>9</sup> - Plafkin et al. 1989X<sup>10</sup> - Popma and Masser 1999X<sup>11</sup> - LTI 2008

**ATTACHMENT C:**

**MATRIX OF PEARSON'S CORRELATION VALUES FOR FISH  
METRICS**

**This page is blank to facilitate double sided printing.**

Pearson Coefficient Matrix

	%DELT_n	%NSCT_n	%NSCT_wt	%INT_n	%INT_wt	%LRN_n	%LRV_wt	%LTHPL_n	%LTHPL_wt	%MOD_n	%MOD_wt	%M_B_OMV	%OMV_n	%OMV_wt	%RRV_n	%RRV_wt	%TC_n
%DELT_n	1	0.265	0.351	0.036	-0.177	-0.053	0.287	-0.010	0.321	0.128	-0.203	0.306	0.313	0.323	0.012	0.280	0.237
%NSCT_n	0.265	1	0.557	0.162	0.144	-0.527	-0.133	-0.527	-0.028	0.336	0.230	-0.385	-0.384	0.005	-0.416	-0.049	0.504
%NSCT_wt	0.351	0.557	1	-0.062	0.000	0.021	0.425	0.022	0.409	0.087	-0.141	0.072	0.078	0.483	0.055	0.434	0.351
%INT_n	0.036	0.162	-0.062	1	0.269	-0.195	-0.249	-0.111	-0.168	0.108	0.111	-0.105	-0.113	-0.171	-0.147	-0.186	0.194
%INT_wt	-0.177	0.144	0.000	0.269	1	0.078	-0.321	0.121	-0.098	-0.077	0.056	-0.362	-0.363	-0.347	0.057	-0.316	-0.002
%LRN_n	-0.053	-0.527	0.021	-0.195	0.078	1	0.582	0.955	0.515	-0.426	-0.494	0.371	0.377	0.407	0.873	0.482	-0.517
%LRV_wt	0.287	-0.133	0.425	-0.249	-0.321	0.582	1	0.563	0.888	-0.167	-0.513	0.518	0.523	0.906	0.574	0.900	-0.185
%LTHPL_n	-0.010	-0.522	0.022	-0.111	0.121	0.955	0.563	1	0.530	-0.471	-0.543	0.328	0.329	0.395	0.828	0.458	-0.485
%LTHPL_wt	0.321	-0.028	0.409	-0.168	-0.098	0.515	0.888	0.530	1	-0.128	-0.485	0.421	0.410	0.808	0.521	0.810	-0.156
%MOD_n	0.128	0.336	0.087	0.108	-0.077	-0.426	-0.167	-0.471	-0.128	1	0.589	0.041	0.026	-0.049	-0.107	-0.024	0.581
%MOD_wt	-0.203	0.230	-0.141	0.111	0.056	-0.494	-0.513	-0.543	-0.485	0.589	1	-0.236	-0.237	-0.407	-0.249	-0.322	0.386
%M_B_OMV_n	0.306	-0.385	0.072	-0.105	-0.362	0.371	0.518	0.328	0.421	0.041	-0.236	1	0.997	0.513	0.375	0.476	-0.099
%OMV_n	0.313	-0.384	0.078	-0.111	-0.363	0.377	0.523	0.329	0.410	0.026	-0.237	0.997	1	0.518	0.376	0.479	-0.090
%OMV_wt	0.323	0.005	0.483	-0.171	-0.347	0.407	0.906	0.395	0.808	-0.049	-0.407	0.513	0.518	1	0.425	0.831	-0.066
%RRV_n	0.012	-0.416	0.055	-0.147	0.057	0.873	0.574	0.826	0.521	-0.107	-0.249	0.375	0.376	0.425	1	0.609	-0.277
%RRV_wt	0.280	-0.049	0.434	-0.186	0.482	0.900	0.458	0.810	-0.024	-0.322	0.476	0.479	0.831	0.609	1	-0.070	0.400
%TC_n	0.237	0.504	0.351	0.194	-0.002	-0.517	-0.185	-0.489	-0.166	0.581	0.386	-0.099	-0.090	-0.066	-0.277	-0.070	1
%TC_wt	-0.352	0.096	-0.101	0.167	0.245	-0.274	-0.536	-0.283	-0.581	0.159	0.539	-0.344	-0.333	-0.523	-0.215	-0.447	0.455
%TOL_n	0.015	-0.230	0.247	-0.132	0.057	0.625	0.499	0.708	0.455	-0.672	-0.591	0.320	0.330	0.434	0.385	0.372	-0.296
%TOL_wt	0.203	0.028	0.466	-0.061	-0.255	0.428	0.726	0.443	0.636	-0.119	-0.374	0.508	0.512	0.740	0.371	0.551	0.040
%CPU	-0.072	0.210	0.068	0.164	0.102	-0.224	-0.064	-0.236	-0.013	0.794	0.485	-0.045	-0.059	0.035	0.025	0.409	0.400
%GEN	0.313	-0.384	0.078	-0.110	-0.363	0.377	0.523	0.329	0.411	0.026	-0.237	0.997	1.000	0.518	0.376	0.480	-0.090
%INT	-0.083	0.117	0.017	0.596	0.420	0.096	-0.099	0.174	0.001	-0.079	-0.013	-0.255	-0.259	-0.094	0.092	-0.082	0.084
%INTOL	0.016	0.125	0.021	0.647	0.223	0.031	-0.043	0.107	0.032	-0.029	0.020	-0.161	-0.168	-0.074	0.044	-0.018	0.130
%LITDT	0.049	0.131	0.066	0.983	0.154	-0.189	-0.198	-0.113	-0.146	0.111	0.094	-0.066	-0.072	-0.124	-0.143	-0.141	0.192
%MDO	-0.114	0.129	0.071	0.041	0.117	-0.035	-0.003	-0.029	0.023	0.570	0.345	-0.032	-0.047	0.073	0.126	0.069	0.251
%NAT	-0.122	0.184	0.183	0.113	0.155	-0.028	0.056	0.033	0.077	0.312	0.180	-0.073	-0.082	0.132	0.068	0.114	0.301
%NFSH	-0.122	0.184	0.183	0.113	0.155	-0.028	0.056	0.033	0.077	0.312	0.180	-0.073	-0.082	0.132	0.068	0.114	0.301
%NMH	-0.275	0.156	0.146	0.014	0.240	0.101	0.088	0.135	0.115	0.077	0.115	-0.174	-0.186	0.071	0.147	0.141	-0.022
%NSUN	0.065	0.152	0.072	0.347	0.000	-0.169	-0.087	-0.116	-0.068	0.473	0.206	0.078	0.064	0.063	-0.056	-0.063	0.474
%M_B_SUN	0.091	0.123	0.029	0.285	-0.038	-0.215	-0.096	-0.166	-0.081	0.544	0.240	0.151	0.135	0.063	-0.073	-0.045	0.404
%PRTOL	0.191	0.089	0.382	-0.294	-0.141	0.154	0.354	0.181	0.297	-0.401	-0.266	0.267	0.285	0.331	0.065	0.317	0.006
%RIV	-0.057	0.099	0.224	0.018	0.100	0.179	0.276	0.230	0.268	0.233	0.000	-0.039	-0.046	0.284	0.270	0.300	0.204
%SA	-0.107	0.183	0.220	0.077	0.186	0.025	0.097	0.095	0.128	0.282	0.100	-0.104	-0.115	0.135	0.096	0.127	0.274
%SUM1	0.091	0.123	0.029	0.285	-0.038	-0.215	-0.096	-0.166	-0.081	0.544	0.240	0.151	0.135	0.063	-0.073	-0.045	0.404
%SUM2	0.065	0.152	0.072	0.347	0.000	-0.169	-0.087	-0.116	-0.068	0.473	0.206	0.078	0.064	0.063	-0.056	-0.063	0.474
%SUM3	0.088	0.134	0.057	0.350	-0.026	-0.175	-0.063	-0.118	-0.047	0.487	0.203	0.100	0.083	0.083	-0.047	-0.019	0.398
%TNI	-0.187	-0.100	0.129	0.016	0.187	0.355	0.337	0.390	0.345	0.093	-0.027	0.107	0.096	0.377	0.434	0.341	-0.072
%TOL	-0.076	0.167	0.264	-0.044	0.110	0.029	0.156	0.107	0.165	0.142	-0.019	-0.073	-0.079	0.200	0.054	0.160	0.241
%WALR	-0.052	0.238	0.057	0.058	0.394	-0.172	-0.151	-0.135	-0.032	0.390	0.411	-0.271	-0.276	-0.101	0.004	-0.058	0.273

NOTE: This matrix does not include: 1. %NRNSCX\_n or %NRNSCX\_wt because there were no data for these metrics (no fish identified in these categories).  
 2. %BIMV, %NSJC, and %SCKR because each of these species metrics had insufficient numbers of species associated with them in the dataset (2 or fewer).



Pearson Coefficient Matrix

	NTC_wrt	NTOL_n	NTOL_wrt	CPUE	GEN	INT	INTOL	LITOT	MOD	NAT	NFSH	NMIN	NSUN	OH_B_SUN	PRTOL	RIV	SR	SUN1
NOELT_n	-0.352	0.015	0.203	-0.072	0.313	-0.083	0.018	0.049	-0.114	-0.122	-0.122	-0.275	0.065	0.091	0.191	-0.057	-0.107	0.091
NOINSC1_n	0.096	-0.230	0.028	0.210	-0.384	0.112	0.125	0.131	0.129	0.184	0.184	0.156	0.152	0.123	0.089	0.099	0.183	0.123
NOINSC2_wrt	-0.101	0.247	0.466	0.068	0.078	0.017	0.021	-0.066	0.071	0.183	0.183	0.146	0.072	0.079	0.382	0.224	0.720	0.029
NOINV_n	0.167	-0.132	-0.061	0.164	-0.110	0.596	0.647	0.983	0.041	0.113	0.113	0.014	0.347	0.285	-0.294	0.018	0.077	0.285
NOINT_wrt	0.245	0.057	-0.255	0.102	-0.363	0.420	0.223	0.154	0.117	0.155	0.155	0.240	0.000	-0.038	-0.141	0.100	0.186	-0.038
NOIRV_n	-0.274	0.625	0.428	-0.224	0.377	0.096	0.031	-0.189	-0.035	-0.028	-0.028	0.101	-0.169	-0.215	0.154	0.179	0.025	-0.215
NOIRV_wrt	-0.538	0.499	0.726	-0.064	0.523	-0.099	-0.043	-0.198	-0.003	0.056	0.056	0.088	-0.087	-0.095	0.354	0.276	0.097	-0.096
NOITNPL_n	-0.282	0.708	0.443	-0.236	0.329	0.174	0.107	-0.113	-0.029	0.033	0.033	0.135	-0.116	-0.166	0.181	0.230	0.096	-0.166
NOITNPL_wrt	-0.581	0.455	0.636	-0.013	0.411	0.001	0.032	-0.146	0.023	0.077	0.077	0.115	-0.068	-0.081	0.297	0.259	0.128	-0.081
NOIMOD_n	0.159	-0.672	-0.113	0.794	0.026	-0.079	-0.029	0.111	0.570	0.312	0.312	0.077	0.473	0.544	-0.401	0.233	0.282	0.544
NOIMOD_wrt	0.539	-0.591	-0.374	0.485	-0.237	-0.013	0.020	0.094	0.345	0.180	0.180	0.115	0.206	0.240	-0.266	0.000	0.100	0.240
NOIH_B_OIMV_n	-0.344	0.320	0.508	-0.045	0.997	-0.255	-0.161	-0.066	-0.032	-0.073	-0.073	-0.174	0.078	0.151	0.267	-0.039	-0.104	0.151
NOIMV_n	-0.333	0.330	0.512	-0.059	1.000	-0.259	-0.168	-0.072	-0.047	-0.082	-0.082	-0.186	0.064	0.135	0.285	-0.046	-0.115	0.135
NOIMV_wrt	-0.523	0.434	0.740	0.035	0.518	-0.094	-0.024	-0.124	0.073	0.132	0.132	0.071	0.063	0.063	0.331	0.284	0.155	0.063
NOIRV_n	-0.215	0.385	0.371	0.025	0.376	0.092	0.044	-0.143	0.126	0.068	0.068	0.147	-0.056	-0.073	0.065	0.270	0.096	-0.073
NOIRV_wrt	-0.447	0.377	0.551	0.025	0.480	-0.082	-0.018	-0.141	0.069	0.114	0.114	0.141	-0.063	-0.045	0.317	0.300	0.127	-0.045
NTC_n	0.455	-0.296	0.040	0.409	-0.090	0.084	0.130	0.192	0.251	0.301	0.301	-0.022	0.474	0.404	0.006	0.204	0.274	0.404
NTC_wrt	1																	
NTOL_n	-0.128	1	0.641	-0.504	0.330	0.061	0.016	-0.131	-0.274	-0.027	-0.027	0.056	-0.182	-0.236	0.625	0.099	0.024	-0.236
NTOL_wrt	-0.153	0.641	1	-0.046	0.512	-0.044	0.000	-0.026	-0.012	0.102	0.102	0.012	0.141	0.096	0.563	0.214	0.142	0.096
CPUE	0.179	-0.504	-0.046	1	-0.060	0.205	0.203	0.137	0.781	0.658	0.658	0.435	0.584	0.634	-0.448	0.567	0.647	0.634
GEN	-0.333	0.330	0.512	-0.060	1	-0.259	-0.168	-0.071	-0.047	-0.082	-0.082	-0.186	0.064	0.135	0.286	-0.047	-0.116	0.135
INT	0.194	0.061	-0.044	0.205	-0.259	1	0.932	0.529	0.156	0.342	0.342	0.218	0.376	0.221	-0.354	0.366	0.369	0.221
INTOL	0.145	0.016	0.000	0.203	-0.168	0.932	1	0.602	0.142	0.348	0.348	0.196	0.448	0.297	-0.332	0.361	0.338	0.297
LITOT	0.134	-0.133	-0.026	0.137	-0.071	0.529	0.602	1	0.001	0.057	0.057	-0.056	0.343	0.280	-0.280	-0.009	0.022	0.280
MOD	0.167	-0.274	-0.012	0.781	-0.047	0.156	0.142	0.001	1	0.775	0.775	0.567	0.544	0.571	-0.485	0.629	0.732	0.571
NAT	0.260	-0.027	0.102	0.658	-0.082	0.342	0.348	0.057	0.775	1	1.000	0.747	0.670	0.642	-0.172	0.869	0.967	0.642
NFSH	0.260	-0.027	0.102	0.658	-0.082	0.342	0.348	0.057	0.775	1.000	1	0.747	0.670	0.642	-0.172	0.869	0.967	0.642
NMIN	0.068	0.056	0.012	0.435	-0.186	0.218	0.196	-0.056	0.567	0.747	0.747	1	0.170	0.167	-0.085	0.687	0.770	0.167
NSUN	0.302	-0.182	0.141	0.584	0.064	0.376	0.448	0.343	0.544	0.670	0.670	0.170	1	0.940	-0.293	0.563	0.645	0.940
OH_B_SUN	0.224	-0.236	0.096	0.684	0.135	0.223	0.297	0.280	0.571	0.642	0.642	0.167	0.940	1	-0.284	0.527	0.606	1.000
PRTOL	0.002	0.625	0.563	-0.448	0.286	-0.354	-0.332	-0.280	-0.495	-0.172	-0.172	-0.085	-0.293	-0.284	1	-0.099	-0.139	-0.284
RIV	0.171	0.099	0.214	0.567	-0.047	0.366	0.361	-0.008	0.629	0.869	0.869	0.687	0.563	0.527	-0.099	1	0.901	0.527
SR	0.216	0.024	0.142	0.647	-0.116	0.369	0.338	0.022	0.752	0.967	0.967	0.730	0.643	0.606	-0.139	0.901	1	0.606
SUN1	0.224	-0.236	0.096	0.684	0.135	0.221	0.297	0.280	0.571	0.642	0.642	0.187	0.940	1.000	-0.284	0.527	0.606	1
SUN2	0.302	-0.182	0.141	0.584	0.064	0.376	0.448	0.343	0.544	0.670	0.670	0.170	1.000	0.940	-0.293	0.563	0.645	0.940
SUN3	0.272	-0.206	0.110	0.609	0.083	0.368	0.449	0.349	0.549	0.658	0.658	0.174	0.970	0.970	-0.323	0.573	0.632	0.970
TN1	0.020	0.230	0.337	0.567	0.095	0.285	0.234	-0.019	0.578	0.730	0.730	0.612	0.427	0.434	-0.041	0.733	0.765	0.434
TOL	0.177	0.146	0.110	0.484	-0.080	0.216	0.195	-0.085	0.538	0.895	0.895	0.679	0.553	0.521	0.113	0.865	0.945	0.521
WPUE	0.240	-0.296	-0.154	0.692	-0.277	0.356	0.251	-0.018	0.612	0.588	0.588	0.461	0.318	0.325	-0.274	0.488	0.609	0.325

**Pearson Coefficient Matrix**

	SUN2	SUN3	TMI	TOL	WPUE
NOELT_(n)	0.065	0.088	-0.187	-0.076	-0.052
NOVSCY_(n)	0.152	0.134	-0.100	0.167	0.238
NOVSCY_(wr)	0.072	0.057	0.129	0.264	0.057
NOVNT_(n)	0.347	0.350	0.016	-0.044	0.058
NOVNT_(wr)	0.000	-0.026	0.187	0.110	0.394
NOVRV_(n)	-0.169	-0.175	0.356	0.029	-0.172
NOVRV_(wr)	-0.087	-0.063	0.337	0.156	-0.151
NOVTHP_(n)	-0.116	-0.118	0.390	0.107	-0.135
NOVTHP_(wr)	-0.068	-0.047	0.345	0.165	-0.032
NOVMOO_(n)	0.473	0.487	0.093	0.142	0.390
NOVMOO_(wr)	0.206	0.203	-0.027	-0.019	0.411
NOVH_B_OMV_(n)	0.078	0.100	0.107	-0.073	-0.271
NOVOMV_(n)	0.064	0.083	0.096	-0.079	-0.276
NOVOMV_(wr)	0.063	0.083	0.377	0.200	-0.101
NOVRV_(n)	-0.056	-0.047	0.434	0.054	0.004
NOVRV_(wr)	-0.063	-0.019	0.341	0.160	-0.098
NOVTC_(n)	0.474	0.398	-0.077	0.241	0.273
NOVTC_(wr)	0.307	0.222	0.020	0.177	0.240
NOVTOL_(n)	-0.182	-0.206	0.230	0.146	-0.296
NOVTOL_(wr)	0.141	0.110	0.337	0.210	-0.154
NOVUE	0.584	0.609	0.567	0.484	0.693
NOVEN	0.064	0.083	0.095	-0.080	-0.277
NOVNT	0.376	0.368	0.285	0.216	0.356
NOVTOL	0.448	0.449	0.234	0.195	0.251
NOVTTOT	0.343	0.349	-0.019	-0.086	-0.018
NOVMO	0.544	0.549	0.578	0.538	0.612
NOVAT	0.670	0.658	0.730	0.895	0.588
NOVFSH	0.670	0.658	0.730	0.895	0.588
NOVWH	0.170	0.174	0.612	0.679	0.461
NOVSUN	1.000	0.970	0.427	0.553	0.318
NOVH_B_SUN	0.940	0.970	0.434	0.521	0.325
NOVPTOL	-0.293	-0.323	-0.041	0.113	-0.274
NOVAV	0.563	0.573	0.733	0.865	0.488
NOVSR	0.645	0.632	0.765	0.945	0.609
SUN1	0.940	0.970	0.434	0.521	0.325
SUN2	1	0.970	0.427	0.553	0.318
SUN3	0.970	1	0.445	0.534	0.327
TMI	0.427	0.445	1	0.721	0.511
TOL	0.553	0.534	0.721	1	0.477
WPUE	0.318	0.327	0.511	0.477	1

**APPENDIX B:**

**TECHNICAL MEMORANDA DESCRIBING  
MACROINVERTEBRATE DATA (FROM BAETIS, INC.)**

This page is blank to facilitate double sided printing

**Technical Memorandum No.1**

**CHARACTERIZATION OF THE MACROINVERTEBRATE COMMUNITY**

**CHICAGO AREA WATERWAY SYSTEM**

**HABITAT RESTORATION EVALUATION AND IMPROVEMENT STUDY**

**Prepared by**

**Baetis Environmental Services, Inc.**

**Chicago, Illinois**

**For**

**LimnoTech, Inc.**

**Ann Arbor, Michigan**

**In support of**

**Metropolitan Water Reclamation District of Greater Chicago**

**Chicago, Illinois**

**February, 2009**

Table of Contents

Summary and Conclusions .....	1
Background .....	2
Methodology .....	2
Macroinvertebrate Community Composition in the CAWS .....	4
General .....	4
Assessment By Sampling Station.....	5
Assessment By Reach .....	8
Comparison Between Sampling Protocols.....	10
Comparison of the Benthic Community Upstream and Downstream of WRPs .....	19
References .....	21

## Summary and Conclusions

A seven-year macroinvertebrate database was developed by the Metropolitan Water Reclamation District of Greater Chicago (District) and used herein to characterize the benthic community within the Chicago Area Waterway System (CAWS). This technical memorandum looked at the macroinvertebrate data combined for the entire CAWS, and separately by AWQM station and by reach. Regardless of whether the data were assessed by the CAWS, by station, or by reach, the results are similar; the macroinvertebrate community is dominated by a few opportunistic Diptera and non-insect taxa.

Nearly half of the taxa collected in the CAWS are from the order Diptera, and almost all are in the family Chironomidae. By abundance, oligochaetes (Phylum Annelida) dominate the benthic community, comprising over 74 percent of all macroinvertebrates collected from the CAWS over the seven-year period. Two species of non-native bivalve, the zebra mussel, *Dreissena polymorpha*, and the closely related Quagga mussel, *Dreissena rostriformis bugensis* comprise 15 percent of the samples. These mussels were collected in very high densities in the Calumet area.

Taxa representing the classic pollution-intolerant organisms, the Ephemeroptera, Plecoptera, and Tricoptera (EPT), are exceedingly scarce in the CAWS. Plecopterans are not present at all, and ephemeropterans and tricopteran are found in very low densities with only a few taxa. At most AWQM stations two or fewer EPT taxa were collected, with EPT densities less than one percent.

An analysis of the differences between sampling methods, i.e. grab samples (ponar) and artificial substrate samples (hester-dendy), show that richness measures (total richness, EPT richness, and diptera richness) are higher in the hester-dendy samples. In contrast, EPT taxa were nearly absent from the ponar collections with EPT richness values of zero for most ponar samples. Clearly, the two sampling methods collected different organisms and in different quantities. The ponar grab samples are heavily dominated by oligochaetes, comprising nearly 100 percent of the samples at many stations (and reaches). While the hester-dendy samples also have high numbers of oligochaetes they comprise far less of the sample than in the ponar samples. At several AWQM stations in the Calumet area the hester-dendy samples had high number of zebra and quagga mussels and lower taxa richness. It is likely that these mussels attached themselves to the hester-dendy artificial substrate, covering the samplers in such high numbers that very few other macroinvertebrates could colonize the sampling apparatus.

We also examined the effect of the District's water reclamation plants (WRP) on



macroinvertebrate communities. We tested the equality of medians for 23 metrics upstream and downstream of the three major treatment plants discharging to the CAWS. We concluded that, for most metrics, there was no difference between the median macroinvertebrate communities upstream and downstream of the three wastewater treatment plants.

## **Background**

Under contract to LimnoTech, Inc., Baetis Environmental Services, Inc. (Baetis) has been retained to analyze macroinvertebrate data collected from the Chicago Area Waterway System (CAWS) between 2001 and 2007. The analysis supports the CAWS Habitat Evaluation and Improvement Study sponsored by the Metropolitan Water Reclamation District of Greater Chicago (MWRDGC). This technical memorandum is an interim deliverable, providing:

- A general description of the macroinvertebrate populations and communities of the CAWS,
- An analysis of any differences that exist in the macroinvertebrate community between sampling stations and reaches, and
- An analysis of any differences that exist between the grab samples (ponar) and artificial substrate samples (hester-dendy).

## **Methodology**

Macroinvertebrates were collected annually each summer from the CAWS from 2001-2007 by MWRDGC, with enumeration and identification by EA Engineering, Science, and Technology, Inc (EA) of Deerfield, IL. For purposes of study, the CAWS has been divided into twenty reaches. Of these twenty, macroinvertebrate data were collected from seventeen reaches (macroinvertebrate data were not collected from reaches 5, 16, and 20). Twenty-three sampling stations are located throughout the seventeen CAWS reaches. Figure 1 shows the locations of the sampling stations and reaches. The District uses both hester-dendy samplers (multi-plate apparati) and ponar dredge samplers at each AWQM station. Most macroinvertebrates were identified to genus; where possible species-level identifications were completed. A detailed description of the methodology is provided by EA in their 2006 report (EA 2006). LimnoTech, Inc. compiled EA's datasets into one database for this project. Descriptive and inferential statistics were derived for the 2001-2007 macroinvertebrate database using SAS software (Vers. 9.1, SAS Institute Inc. Cary, N.C.)

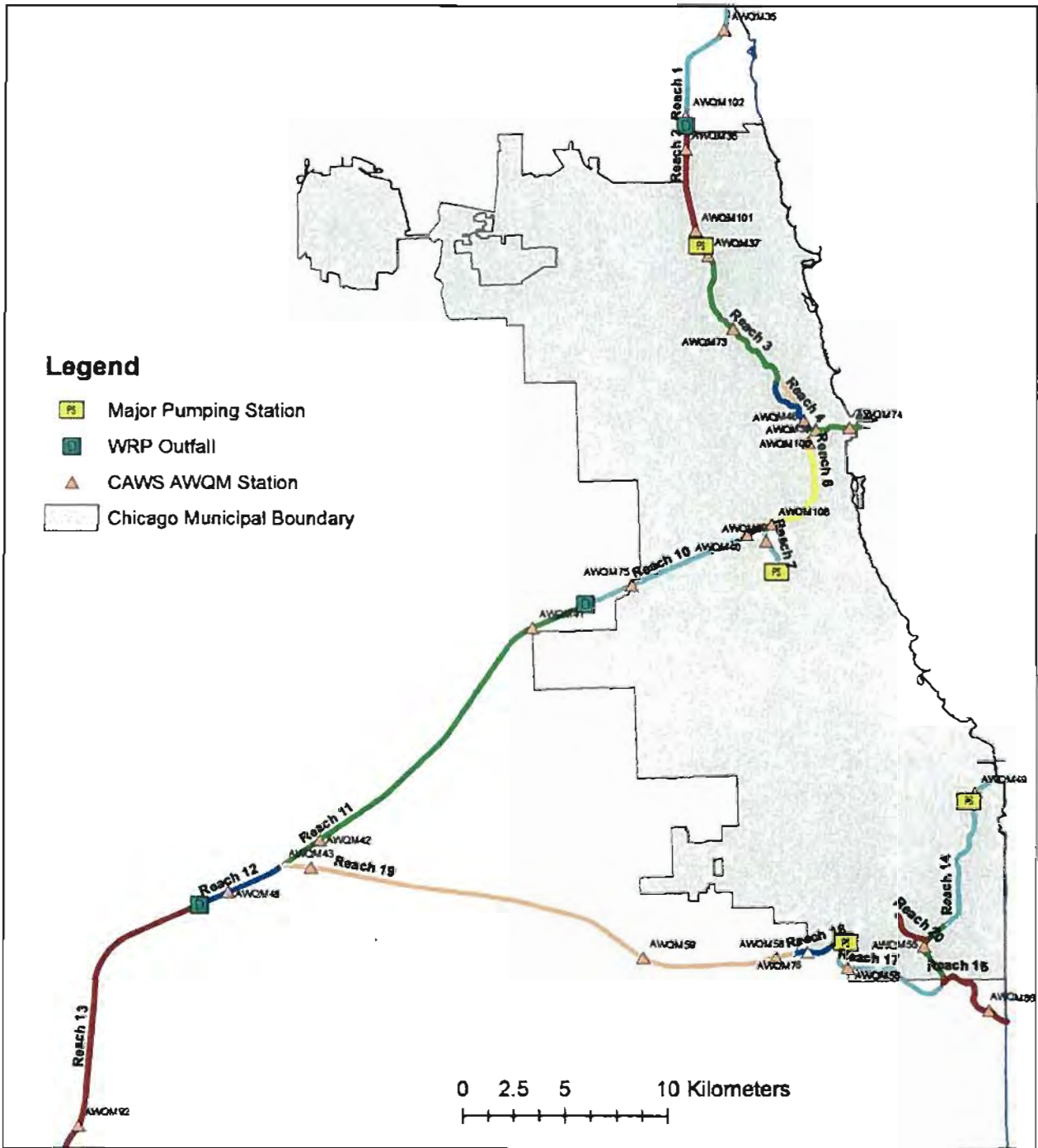


Figure 1. AWQM Station and Reach Designations

## Macroinvertebrate Community Composition in the CAWS

### General

Over eight million macroinvertebrates were collected and identified between 2001 and 2007. They represented 130 macroinvertebrate taxa, though nearly all the diversity can be attributed to the order Diptera (true flies) and to non-insect taxa such as Oligochaeta, flatworms, leeches, isopods, amphipods, snails, and bivalves (Table 1). Nearly half of the taxa (63) were from the order Diptera, almost all within the family Chironomidae, a family of non-biting flies that can often comprise at least fifty percent of the species diversity in a stream (Coffman et al. 1996). Forty-four non-insect taxa were collected from the CAWS. Outside of the family Chironomidae, taxa richness of the insect community within the CAWS was low. The pollution-sensitive orders Ephemeroptera (mayflies), Trichoptera (caddisflies), and Plecoptera (stoneflies) (EPT) were poorly represented; only fourteen taxa within these orders were collected.

The macroinvertebrate community of the CAWS is dominated by a few pollution-tolerant taxa. Oligochaetes, a class of pollution-tolerant aquatic worms found in soft mud bottoms, comprised nearly 73 percent of all macroinvertebrates collected from the CAWS (Table 2). Two species of non-native bivalve, the zebra mussel, *Dreissena polymorpha*, and the closely related Quagga mussel, *Dreissena rostriformis bugensis* comprised 15 percent of the samples. These invasive species were introduced into the Great Lakes region in ballast water from oceangoing vessels and have had far-reaching and deleterious impacts (Smith 2001, USGS 2008, USDA 2008). True flies (Order Diptera) are the third most abundant taxon, at nearly 6 percent of the collections. Within this order, the family Chironomidae, a family often associated with environmental perturbation, accounted for nearly all the diptera present. In comparison, the densities of pollution-sensitive mayflies, caddisflies, and stoneflies that were collected were very low, comprising only 0.001 percent of the samples. These taxa are often the first to decline in a stressed system.

A shift towards dominance by a few taxa indicates environmental stress. In healthy, natural aquatic systems the macroinvertebrate community is not dominated by a few taxa but, instead, has a more balanced distribution. The percent contribution of such organisms as Oligochaeta and Diptera are expected to increase in response to stream perturbation. These dominant taxa collected from the CAWS are opportunistic taxa that can exist in stressed or man-made environments and are often indicators of poor water quality, poor sediment quality, and/or poor

habitat quality.

### **Assessment By Sampling Station**

A description of the macroinvertebrate community collected from each sampling station is provided below. In general, while there are some notable differences between stations, the data show that all stations support a macroinvertebrate community dominated by a few opportunistic taxa in the Diptera and non-insect groups. Figure 1 shows the AWQM stations that the District samples in the CAWS.

Table 1 provides counts of total taxa collected from each station. The highest total richness values were found at AWQM 92 (58 taxa) and AWQM 76 (54 taxa). These stations had more samples taken (28) than many other stations; thus the higher richness values may be a result of increased sampling effort rather than a larger 'pool' of macroinvertebrates. Lowest total richness was found at AWQM 99 (14 taxa) and at AWQM 40 (19 taxa), two of the least sampled stations.

EPT richness was low for all stations. In general, two or fewer EPT taxa were collected from each station, although there were some exceptions. AWQM 92 and AWQM 75 had the highest EPT richness values with 7 taxa (AWQM 92) and 6 taxa (AWQM 75). Again, AWQM 92 was one of the most sampled stations; AWQM 75 was also sampled more than many stations. Even considering the number of samples taken at these stations, EPT richness values were low. AWQM stations 46, 99, and 101 had EPT richness values of zero. AWQM 46 was also one of the most sampled stations so an EPT richness value of zero certainly indicates poor aquatic conditions at this site.

Table 2 provides a comparison by station of the macroinvertebrate community composition and functional feeding groups. By abundance, oligochaetes dominate the macroinvertebrate community at most stations. Oligochaetes were found in the highest densities, comprising over half the macroinvertebrates in samples from all but three stations. In fifteen of the 23 stations, oligochaetes comprised over 70 percent or more of the samples. There were only three stations (AWQM 49, 55, and 56) where oligochaetes represented less than half the macroinvertebrates within each sample. Samples from these three stations contained large numbers of zebra mussels and quagga mussels, particularly AWQM 55 (94 percent of the sample) and AWQM 56 (50 percent of the sample).

Table 1  
TAXA RICHNESS BY SAMPLING STATION IN THE CHICAGO AREA WATERWAY SYSTEM

	CAWS	AWQM Sampling Station																						
		35	36	37	39	40	41	43	46	49	55	56	58	59	73	74	75	76	92	99	100	101	102	108
No. of Samples	8	28	8	8	8	28	8	28	8	26	8	8	28	8	8	27	28	28	8	8	8	8	8	8
Total Richness	130	43	45	24	30	19	41	39	39	49	46	30	36	48	28	36	40	54	58	14	32	22	31	28
EPT Richness	14	2	4	2	1	1	1	3	0	4	3	2	1	2	2	2	6	3	7	0	1	0	2	2
Diptera Richness	63	26	24	14	14	7	19	19	17	30	27	20	16	25	14	23	15	23	25	7	20	11	20	10
Non-insect Richness*	44	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

\*Calculated for the CAWS.

Table 2  
MACROINVERTEBRATE COMMUNITY COMPOSITION AND TROPHIC STRUCTURE METRICS BY SAMPLING STATION, IN THE CHICAGO AREA WATERWAY SYSTEM

	CAWS	AWQM Sampling Station																						
		35	36	37	39	40	41	43	46	49	55	56	58	59	73	74	75	76	92	99	100	101	102	108
% Diptera	5.9	9.2	7.8	1.6	16.9	4.0	4.7	19.5	4.5	8.1	0.5	11.1	4.2	21.3	3.2	2.5	8.3	9.9	3.5	30.0	8.7	6.8	4.4	16.5
% Chironomid	5.9	9.2	7.8	1.6	16.9	4.0	4.7	19.5	4.5	8.1	0.5	11.1	4.2	21.3	3.2	2.5	8.3	9.9	3.5	30.0	8.7	6.8	4.4	16.5
% Oligochaeta	73	89.4	86.2	95.3	58.0	92.3	87.7	71.9	90.9	15.7	2.9	27.3	93.3	59.9	95.1	72.9	82.5	55.8	90.4	68.4	88.1	81.0	95.1	64.8
% Dreissena sp.	15	0.08	0.01	0	11.7	0	0.38	0.2	0.005	23.6	94.1	50.0	0.9	11.3	0	19.8	0	20.8	0.02	0	0.24	0.04	0	0.45
% EPT	0.001	0.01	0.01	0.003	0.01	0.55	0.12	2.12	0	0.13	0.08	2.24	0.01	0.18	0.002	0.07	0.04	0.27	0.07	0	0.02	0	0.01	1.6
% Shredders	-	0.9	3.3	0.1	0.61	0.21	0.1	7.4	0.2	3.7	0.12	2.7	0.22	1.5	0.29	1.2	0.03	1.36	0.05	0.13	3.0	0.15	0.1	0.45
% Scrapers	-	0.15	0.06	0.02	2.6	0.01	0.09	0.08	0.03	0.16	0.08	0.01	0.33	0.89	0.02	0.17	0.12	0.98	0.84	1.2	0.65	0.1	0.03	0.5
% Collector-filterers	-	0.52	0.01	0.01	11.8	0.21	0.43	0.44	0.06	23.8	94.2	50.1	0.98	11.8	0.003	19.8	0.33	21.3	0.44	0.11	0.31	0.06	0.01	2.5
% Collector-gatherers	-	96.4	93.5	96.4	74.2	96.1	92.2	77.3	95.1	18.9	3.3	36.3	96.4	76.8	97.9	73.9	90.3	62.7	92.8	97.9	95.9	87.6	99.0	79.9
% Predators	-	1.5	5.5	3.3	5.4	3.1	6.4	14.4	4.3	52.6	0.5	12.3	1.7	6.8	1.9	0.57	8.7	10.0	3.9	0.82	0.94	11.5	0.75	14.7

The invasive zebra mussel and the quagga mussels appear to have a patchy distribution within the CAWS with the highest numbers found in AWQM 55 (94 percent) followed by AWQM stations 56, 49 and 76 (see Table 2 - % *Dreissena* sp.). These stations are in the Calumet area, an area that supports heavier barge traffic than the other reaches on the CAWS. It is probable that barge and boat traffic in this area contributed to the spread of zebra and quagga mussels in this area, although we cannot dismiss the Lake Michigan diversion flows through the Calumet River. Far fewer numbers of these species are found at other AWQM stations in the CAWS, and the mussels are absent from many other reaches.

The average percent EPT (PER\_EPT) was very low for all stations with the highest percentages just at 2 percent (AWQM 43 and AWQM 56) (Table 2). AWQM 108 had the third highest PER\_EPT at just over 1.5 percent. The remaining stations had average EPT densities of less than 1 percent per sample.

In non-wadeable natural rivers the typical macroinvertebrate assemblage is dominated by collector functional feeding groups (USEPA 2006). In the CAWS, nearly all stations are dominated by the collector functional feeding group. At many stations collector-gatherers, heavily represented by oligochaetes, comprise 90+ percent of the community. These taxa feed by collecting organic particles from the debris and sediments on the bed of a stream. High numbers of collector filterers are found at only a few stations, AWQM 55 (94 percent), AWQM 56 (50 percent) and AWQM 49 (24 percent). Collector-filterers feed by collecting organic particles from the water column using a variety of filters. Zebra mussels and quagga mussels are present in AWQM stations 55 and 56 in very high numbers; these collector-filterer taxa also make up a large part of the macroinvertebrate community collected in AWQM 49 and AWQM 76 although in smaller numbers. From the data, it is evident that the relative abundance of the different functional feeding groups is closely correlated with the relative abundance of oligochaetes, zebra mussels, and quagga mussels.

Shredders appear in the samples in far fewer numbers, comprising less than one percent of the macroinvertebrate population at many stations. AWQM 43, at 7.4 percent, has the highest proportion of shredders. Shredders feed on leaf litter and other organic material from the riparian zone in smaller, natural, headwater streams. They convert this leaf litter, or coarse particulate organic matter (CPOM) to fine particular organic matter (FPOM) which is consumed by the collector functional feeding group in downstream reaches. The CAWS, which is a larger non-wadeable, manmade waterway, supports a macroinvertebrate community that is strongly

comprised of the collector functional feeding group. It appears from the data that the influence of the riparian zone and CPOM input is reduced throughout the waterway and limits this feeding group.

Scrapers are rarer than shredders in the CAWS with the highest percentages collected from AWQM 39 (2.6 percent) and AWQM 99 (1 percent). All other stations had scraper percentages less than one percent.

### **Assessment By Reach**

For planning purposes, the macroinvertebrate metrics were also calculated by reach; the results are shown in Table 3. Designated reaches are shown in Figure 1. The trends observed by reach correspond to the trends observed at each sampling station, i.e. within each reach the macroinvertebrate community is dominated by a few taxa in the Diptera and non-insect groups.

Consistent with the station data, the highest total richness values are found in reaches 13, 19, and 18 with 58 taxa found in reaches 13 and 19, and 54 taxa collected in reach 18. Reaches 13 and 18 contain stations AWQM 92 and 76 which had the highest taxa richness out of all samples; reach 19 contains AWQM 43 and 59, which also had high taxa richness scores. Lowest total richness was found in reaches 12 and 7, with 6 and 14 taxa, respectively. These are also two of the least sampled reaches. Reach 7 is the heavily contaminated Bubbly Creek waterway so it is not surprising that the richness values are low. Reach 12 was sampled only once during the 2001-2007 period.

With the exception of reaches 10 and 13, four or fewer EPT taxa were collected from each reach. Six EPT taxa were collected from reach 10; seven EPT taxa were collected from reach 13. The stations with the highest EPT richness, AWQM 92 and AWQM 75, are the only stations located within these reaches.

In nine of the 17 reaches, oligochaetes comprised 80 percent or more of the samples. There were only four reaches (reaches 12, 14, 15, and 17) where oligochaetes represented less than half the macroinvertebrates within each sample. Reaches 14, 15, and 17 are in the Calumet area where other non-insect invertebrates, primarily invasive zebra and quagga mussels, have replaced oligochaetes as the most abundant organism.

The average percent dominance is also provided in Table 3. As expected, the average percent dominance for each reach is high. Again, the assemblage within each reach is dominated by a



few taxa which have resulted in lowered diversity. With the exception of reach 12 (which was only sampled once), each reach has average percent dominance values over 65 percent; with many reaches with average percent dominance values over 80 percent. Based upon the single sample, reach 12 does not appear to be dominated by oligochaetes nor are there one or two taxa exceedingly dominant in the samples. However, a close look at the macroinvertebrates collected from reach 12 during the single sampling event indicate that a sample dominated by oligochaetes, flatworms, chironomids, leeches, and the exotic Asiatic clam *Corbicula* and zebra mussel. Reaches 14, 15, and 17 have high average percent dominance values, however, the samples collected from these reaches are not dominated by oligochaetes; instead, these communities are dominated by hydra and quagga mussels (reach 14) and quagga mussels and zebra mussels (reaches 15 and 17).

The percent EPT is very low for all reaches, the maximum being 2% (in reach 17). While reach 17 has low EPT richness with only 2 taxa, the numbers of individuals appear to be higher than in other reaches. The remaining reaches have average EPT densities of less than 1 percent per sample.

High numbers of collector filterers, present as zebra mussels, quagga mussels, and the Asiatic clam, *Corbicula fluminea* (reach 12 only), are found in only a few reaches. Reach 12 had 39 percent collector filterers, reach 14 had 24 percent, reach 15 had 94 percent, and reach 17 had 50 percent. AWQM 49, 55, and 56 are the only stations within reaches 14, 15, and 17 so the results are the same for both. Shredders appear throughout in far fewer numbers, comprising less than one percent in many reaches. Reach 14, at 3.7 percent, has the highest number of shredders.

Table 3

## MACROINVERTEBRATE COMMUNITY COMPOSITION AND TROPHIC STRUCTURE METRICS BY REACH

	CAWS	1	2	3	4	6	7	8	9	10	11	12	13	14	15	17	18	19
Total Richness	130	50	47	33	39	45	14	38	19	40	44	6	58	49	46	30	54	58
EPT Richness	14	3	4	2	0	2	0	3	1	6	2	0	7	4	3	2	3	4
Diptera Richness	63	30	24	18	17	29	7	15	7	15	21	1	25	30	27	20	23	28
Non-insect Richness	44	16	18	13	22	14	7	19	10	19	19	5	24	15	16	8	25	24
% Diptera	5.9	5.8	7.7	2.5	4.5	5	30	16.7	4	8.3	4.7	8.7	3.5	8.1	0.5	11.1	9.9	11.1
% Chir	5.9	5.8	7.7	2.5	4.5	5	30	16.7	4	8.3	4.7	8.7	3.5	8.1	0.5	11.1	9.9	11.1
% Olig	73	93.3	85.7	95.2	90.9	79	68.4	60.6	92.3	82.5	87.6	13.1	90.4	15.7	2.9	27.3	55.8	80.5
% Dreis.	15	0.02	0.01	0	0.005	12	0	7	0	0	0.4	35	0.02	24	94	50	21	4
% Dom*	-	76	74.7	81.8	85.6	78.4	86.8	75.0	85.1	85.7	82.2	34.8	69.7	71.2	86.1	74.3	71.0	65.9
% EPT	0.001	0.01	0.01	0.003	0	0.05	0	0.62	0.55	0.04	0.18	0	0.07	0.13	0.08	2.2	0.27	0.27
% Shred	-	0.34	3.01	0.21	0.2	1.9	0.13	0.55	0.21	0.03	0.1	0	0.05	3.7	0.1	2.7	1.4	1.3
% Scrapers	-	0.07	0.06	0.02	0.03	0.37	1.2	1.8	0.01	0.12	0.1	0	0.84	0.16	0.08	0.01	1.0	0.49
% Cltr-fltrs	-	0.16	0.02	0.007	0.06	11.9	0.11	8.2	0.21	0.33	0.43	39.1	0.43	23.8	94.2	50.1	21.3	4.42
% Cltr-gthrs	-	98.2	93.0	97.3	95.1	82.8	97.8	76.4	96.1	90.3	92.1	13.0	92.8	18.9	3.25	36.3	62.7	88.3
% Predtrs	-	1.0	6.1	2.5	4.32	0.72	0.82	9.04	3.12	8.71	6.44	47.8	3.9	52.6	0.5	12.3	10.0	4.52

\*This value represents the average percent dominance per sample.

### **Comparison Between Sampling Protocols**

We investigated differences, if any, between the grab samples (ponar) and artificial substrate samples (hester-dendy). Clearly, the two sampling methods collect different organisms in different quantities. Table 4 compares the results of the two sampling methods for the entire CAWS; Tables 5 through 7 compare the results of the two sampling methods by sampling station. Comparisons by reach are presented in Tables 7 and 8.

Greater numbers of macroinvertebrates were collected using a ponar sampler than the hester-dendy apparatus (Table 4); importantly however, taxa richness was much higher in the hester-dendy samples. Richness measures were, in fact, much higher in the hester-dendy samples for every richness category assessed. Thus, while higher numbers were collected with the ponar in the CAWS, the ponar samples collected fewer taxa and had overall lower diversity.

Community composition measures for the CAWS show that the hester-dendy samples had higher percentages of Diptera and EPT individuals. Hester-dendy samples also had higher numbers of the invasive zebra mussel and quagga mussel (Genus *Dreissena*). Ponar grab samples, in comparison, had very high numbers of oligochaetes; comprising 97 percent of the ponar samples for the CAWS. Oligochaetes make up only 65 percent of the hester-dendy samples for the CAWS (Table 4).

Because oligochaetes, which fall within the collector-gatherer functional feeding group, make up the vast majority of the macroinvertebrate collections in the ponar grab samples, it is not surprising that this sampling technique has a much higher percentage of collector-gatherers. In contrast, the hester-dendy samples have a much lower percentage of collector-gatherers and a higher percentage of the shredder, scraper, collector-filterer, and predator functional feeding groups. This is likely the result of the higher macroinvertebrate diversity found in the hester-dendy samples.

Station-wise and reach-wise comparisons show similar patterns (Tables 5-7 and Tables 8-9). With only two exceptions (AWQM 101, one of two stations in reach 2, and AWQM 55 the only station in reach 15) total richness and EPT richness values are higher in the hester-dendy samples for each station and reach. The ponar grab method did not collect EPT taxa from most stations, while the hester-dendy method collected EPT taxa from all stations with the exception of three. With few exceptions, Diptera richness is also higher in the hester-dendy samples.

**Table 4**

**COMPARISON OF PONAR AND HESTER-DENDY SAMPLING METHODS**

	<b>Ponar</b>	<b>Hester-Dendy</b>
Total # Samples Collected	176	171
Total # of Individuals	5,091,260	3,192,962
<b>Richness Measures</b>		
Total Richness	81	111
EPT Richness	5	13
Ephemeroptera Richness	2	5
Tricoptera Richness	3	8
Diptera Richness	43	53
<b>Community Composition and Functional Feeding Group</b>		
% Diptera	1.9	12
% Chironomidae	1.9	12
% Oligochaeta	97	65
% Dreissena	0.4	39
% EPT	0.005	0.3
% Shredders	0.3	2.5
% Scrapers	0.03	0.6
% Collector-filterers	0.6	38.7
% Collector-gatherers	97.5	47.3
% Predators	1.7	10

As discussed above, throughout the CAWS higher numbers of individuals were collected using the ponar sampling method (Table 4). That said, when looking at a station comparison and a reach by reach comparison, one can see that in approximately half the stations and reaches the hester-dendy samples have higher numbers of macroinvertebrates than the ponar samples. It is interesting to note that in AWQM 55 (the only station in reach 15) macroinvertebrates were collected in vastly greater numbers using the hester-dendy (1,079,540 individuals) than the ponar (39,746 individuals) yet the hester-dendy samples at this site have lower richness values for several metrics in comparison to the ponar samples. The high numbers of macroinvertebrates coupled with low diversity in reach 15 can be explained by the very high numbers of the invasive zebra mussels and quagga mussels that dominate the hester-dendy samples. It is likely that these mussels attached themselves to the hester-dendy artificial substrate in reach 15 (and to a lesser extent in reaches 14, 17, and 18) covering the samplers in such high numbers that very few other

macroinvertebrates could colonize the sampling apparatus.

As in the overall CAWS, the ponar grab samples are heavily dominated by oligochaetes, comprising nearly 100 percent of the samples at many stations (and reaches). While the hester-dendy samples also have high numbers of oligochaetes they comprise far less of the sample than in the ponar samples. In conjunction with the high oligochaete percentages, collector-gatherers are the dominant functional feeding group in the ponar samples collected from each station (and reach).

Table 5

## COMPARISON OF METRICS FOR PONAR AND HESTER-DENDY SAMPLING METHODS, AWQM STATIONS 35-43

	AWQM 35		AWQM 36		AWQM 37		AWQM 39		AWQM 40		AWQM 41		AWQM 43	
	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD
Total # Samples	4	4	14	14	4	4	4	4	4	4	14	14	4	4
Total # of Individuals (m <sup>2</sup> )	153,896	9,559	1,441,758	487,492	275,037	46,499	10,794	25,059	54,743	16,164	429,809	362,938	53,250	18,059
Total Richness	19	37	27	39	11	21	11	27	7	18	23	33	10	35
EPT Richness	0	2	0	4	0	2	0	1	0	1	1	1	0	3
Diptera Richness	12	21	16	19	8	11	5	13	4	6	11	13	6	16
% Diptera	6.7	48.7	1.6	26.2	1.2	3.7	1.1	23.7	0.4	16.0	0.7	9.4	10.0	47.4
% Chironomidae	6.7	48.7	1.6	26.2	1.2	3.7	1.1	23.7	0.4	16.0	0.7	9.4	10.0	47.4
% Oligochaeta	92.6	37.6	97.4	53.2	98.6	75.8	59.2	57.5	99.3	68.5	96.2	77.6	89.7	19.6
% Dricusena sp.	6.7	0.11	1.6	0.03	1.2	0	1.1	0.29	0.39	0	0.74	0.82	10.0	0.77
% EPT	0	0.11	0	0.05	0	0.02	0	0.01	0	2.4	0.003	0.26	0	8.4
% Shredders	0.88	1.2	0.17	12.6	0.08	0.2	0.13	0.82	0.26	0.04	0.08	0.13	0	29.1
% Scrapers	0.04	2.0	0.005	0.22	0	0.16	0	3.7	0	0.04	0.11	0.07	0	0.30
% Collector-filterers	0.54	0.11	0	0.05	0.01	0.02	38.6	0.29	0.26	0.02	0.01	0.92	0.19	1.7
% Collector-gatherers	97.3	81.9	98.6	78.6	99.3	79.3	59.8	80.4	99.6	84.3	96.8	86.7	89.8	40.4
% Predators	1.1	7.4	2.0	15.8	0.76	18.4	1.3	7.2	0.16	13.1	2.8	10.6	9.9	27.6

Table 6

## COMPARISON OF METRICS FOR PONAR AND HESTER-DENDY SAMPLING METHODS, AWQM STATIONS 46-73

	AWQM 46		AWQM 49		AWQM 55		AWQM 56		AWQM 58		AWQM 59		AWQM 73	
	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD
Total # Samples	14	14	4	4	14	12	4	4	4	4	14	14	4	4
Total # of Individuals (m2)	213,764	186,046	11,942	36,118	39,746	1,079,540	12,301	47,485	407,934	31,026	118,501	124,202	404,117	42,874
Total Richness	21	32	29	36	36	27	13	25	6	33	25	43	10	22
EPT Richness	0	0	0	4	2	1	0	2	0	1	0	2	0	2
Diptera Richness	10	15	24	18	20	15	10	15	2	15	13	21	5	9
% Diptera	0.4	9.3	27.2	1.9	4.9	0.34	11.2	11.1	0.82	48.0	6.1	35.8	0.51	28.6
% Chironomidae	0.4	9.3	27.2	1.9	4.9	0.34	11.2	11.1	0.82	48.0	6.1	35.8	0.51	28.6
% Oligochaeta	99.4	81.0	55.5	2.6	63.4	0.71	88.6	11.4	99.0	18.4	92.1	29.1	99.4	54.0
% Dreissena sp	0.40	0.002	27.2	25.8	4.9	96.5	11.2	62.9	0	12.7	0.59	21.4	0	0
% EPT	0	0	0	0.17	0.14	0.07	0	2.8	0	0.13	0	0.36	0	0.03
% Shredders	0.03	0.39	12.1	0.97	1.9	0.04	1.3	3.1	0	3.2	0.07	2.9	0	3.0
% Scrapers	0	0.07	0	0.22	0.18	0.08	0	0.02	0	4.7	0.01	1.7	0	0.16
% Collector-filterers	0.09	0.01	17.1	26.0	30.0	96.5	0.23	63.0	0.08	12.8	1.48	21.7	0	0.03
% Collector-gatherers	99.6	90.0	66.1	3.2	64.6	0.99	91.1	22.1	99.0	62.7	92.7	61.6	99.7	80.8
% Predators	0.38	8.9	7.7	67.5	2.2	0.44	7.4	13.6	0.82	12.9	5.38	8.1	0.49	15.4

Table 7

## COMPARISON OF METRICS FOR PONAR AND HESTER-DENOY SAMPLING METHODS, STATIONS 74-108

	AWQM 74		AWQM 75		AWQM 76		AWQM 92		AWQM 99		AWQM 100		AWQM 101		AWQM 102		AWQM 108	
	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD
Total # Samples	4	4	14	13	14	14	14	14	4	4	4	4	4	4	4	4	4	4
Total # of Individuals (m2)	19,148	7,711	41,841	158,637	317,984	231,845	612,583	95,939	7,005	25,763	7,018	11,260	124,013	77,772	318,945	53,659	10,493	12,451
Total Richness	8	36	14	36	34	47	17	52	3	13	5	29	17	16	19	22	10	25
EPT Richness	0	2	1	5	0	3	1	7	0	0	0	1	0	0	0	2	0	2
Diptera Richness	5	23	7	13	17	19	9	20	1	6	2	19	10	7	11	14	5	9
% Diptera	0.98	6.4	0.82	10.2	5.2	16.4	1.4	16.9	0.82	37.9	0.41	13.8	1.3	15.5	0.46	27.6	3.3	27.6
% Chironomidae	0.98	6.4	0.82	10.2	5.2	16.4	1.4	16.9	0.82	37.9	0.41	13.8	1.3	15.5	0.46	27.6	3.3	27.6
% Oligochaeta	97.2	12.7	97.8	78.4	90.6	8.1	97.7	43.6	99.0	60.1	99.2	81.1	97.9	54.2	99.4	69.6	92.2	41.7
% Dreissena sp.	1.9	64.2	0	0	0.28	49.0	0	0.14	0	0	0.21	0.25	0.06	0	0	0	0.96	0.03
% EPT	0	0.23	0.03	0.04	0	0.64	0.02	0.35	0	0	0	0.03	0	0	0	0.07	50	2.9
% Shredders	0.45	3.1	0.07	0.02	1.81	0.75	0	0.37	0	0.17	0.21	4.7	0.03	0.32	0.01	0.64	0.14	0.72
% Scrapers	0	0.61	0.03	0.14	0.07	2.2	0.07	5.8	0	1.5	0	1.1	0	0.25	0	0.2	0	0.98
% Collector-filterers	1.87	64.3	1.0	0.14	1.1	49.1	0.32	1.2	0	0.14	0.41	0.25	0.09	0	0.009	0	4.5	0.86
% Collector-gabblers	97.5	15.4	98.0	88.3	91.3	23.4	98.0	59.2	99.0	97.6	99.2	93.8	99.1	69.3	99.5	96.3	93.0	68.8
% Predators	0.23	1.4	0.76	10.8	2.9	19.9	1.2	21.1	1.0	0.77	0	1.5	1.0	28.3	0.43	2.6	2.3	25.1



Table 8  
COMPARISON OF METRICS FOR PONAR AND HESTER-DENDY SAMPLING METHODS, REACHES 1-10,

	Reach 1		Reach 2		Reach 3		Reach 4		Reach 6		Reach 7		Reach 8		Reach 9		Reach 10	
	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD
Total # Samples Collected	8	8	18	18	8	8	14	14	8	8	4	4	8	8	4	4	14	13
Total # of Individuals (m <sup>2</sup> )	472,840	63,218	1,565,771	565,264	679,154	89,373	213,764	186,046	26,166	18,971	7,005	25,763	21,286	37,510	54,743	16,165	41,841	158,637
Total Richness	25	42	30	41	16	29	21	32	11	44	3	13	15	34	7	18	14	36
EPT Richness	0	3	0	4	0	2	0	0	0	2	0	0	0	3	0	1	1	5
Ephemeroptera Richness	0	2	0	1	0	1	0	0	0	0	0	0	0	1	0	0	1	2
Tricoptera Richness	0	1	0	3	0	1	0	0	0	2	0	0	0	2	0	1	0	3
Diptera Richness	15	25	17	20	10	14	10	15	7	28	1	6	6	14	4	6	7	13
% Diptera	2.5	31	1.5	25	0.8	16	0.4	9	0.8	11	0.8	38	2.2	25	0.4	16	0.8	10
% Chironomidae	2.5	31	1.5	25	0.8	16	0.4	9	0.8	11	0.8	38	2.2	25	0.4	16	0.8	10
% Oligochaeta	97	65	97	53	99	65	99	81	98	53	99	60	75	52	99	68	98	78
% Dreissena sp.	0.02	0.02	0.005	0.03	0	0	0.007	0	1.4	26	0	0	20	0.2	0	0	0	0
% EPT	0	0.07	0	0.04	0	0.02	0	0	0	0.11	0	0	0	0.97	0	2.4	0	0.04
% Shredders	0.29	0.7	0.16	10.9	0.03	1.6	0.03	0.4	0.38	4.1	0	0.17	0.14	0.79	0.26	0.04	0.07	0.03
% Scrapers	0.01	0.5	0.005	0.23	0	0.16	0	0.07	0	0.87	0	1.53	0	2.8	0	0.05	0.03	0.14
% Collector-filterers	0.18	0.02	0.01	0.05	0	0.03	0.09	0.01	1.48	26.3	0	0.14	21.78	0.48	0.26	0.02	1.03	0.14
% Collector-gatherers	98.8	94.1	98.6	77.3	99.6	80	99.6	90	97.9	61.9	99	97.6	76.2	76.3	99.6	84.3	98	88
% Predators	0.65	3.4	1.93	17.5	0.6	16.9	0.38	8.9	0.17	1.5	1.02	0.77	1.82	13.1	0.16	13.1	0.76	10.8

PN = Ponar Grab Sample

HD = Hester-dendy Artificial Substrate Sample

Table 9

## COMPARISON OF METRICS FOR PONAR AND HESTER-DENDY SAMPLING METHODS, REACHES 11-20

	Reach 11		Reach 12		Reach 13		Reach 14		Reach 15		Reach 17		Reach 18		Reach 19	
	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD	PN	HD
Total # Samples Collected	17	16	1	0	14	14	4	4	14	12	4	4	14	14	22	22
Total # of Individuals (m <sup>2</sup> )	434,115	367,801	330	-	612,583	95,940	11,942	36,118	39,746	1,079,540	12,301	47,485	317,984	231,845	579,687	173,287
Total Richness	26	35	6	-	17	52	29	36	36	27	13	25	34	47	28	52
EPT Richness	1	2	0	-	1	7	0	4	2	1	0	2	0	3	0	4
Ephemeroptera Richness	0	0	0	-	0	2	0	0	1	0	0	1	0	0	0	2
Tricoptera Richness	1	2	0	-	1	5	0	4	1	1	0	1	0	3	0	2
Diptera Richness	12	14	1	-	9	20	24	18	20	15	10	15	17	19	15	23
% Diptera	0.7	9	8.7	-	1.4	17	27.2	2	4.9	0.34	11.2	11	5.2	16	2.7	39
%Chironomidae	0.7	9	8.7	-	1.4	17	27.2	2	4.9	0.34	11.2	11	5.2	16	2.7	39
% Oligochaeta	96	77	13	-	98	44	56	3	63	1	89	11	91	8	97	26
% Dreissena sp.	0	0.8	35	-	0	0.14	17	26	30	97	0.2	63	0.3	49	0.12	18
% EPT	0.003	0.39	0	-	0.023	0.35	0	0.17	0.145	0.07	0	2.8	0	0.64	0	1.1
% Shredders	0.08	0.13	0	-	0	0.37	12.14	0.97	1.91	0.04	1.28	3.1	1.81	0.75	0.01	5.6
% Scrapers	0.11	0.09	0	-	0.07	5.8	0	0.22	0.18	0.08	0	0.01	0.07	2.2	0.002	2.1
% Collector-filterers	0.01	0.93	19.13	-	0.32	1.2	17.1	26	30.1	96.5	0.23	63	1.05	49	0.38	18
% Collector-gatherers	96.9	86.6	13.1	-	98	59.2	66.1	3.2	64.6	0.99	91.1	22.1	91.3	23.4	96.8	59.6
% Predators	2.7	10.8	47.8	-	1.2	21	7.7	67.5	2.2	0.44	7.3	13.6	2.8	19.9	2.6	11

PN = Ponar Grab Sample

HD = Hester-dendy Artificial Substrate Sample

### **Comparison of the Benthic Community Upstream and Downstream of WRPs**

We examined the effect of sampling method on measuring the effects of District water reclamation plants (WRP) on macroinvertebrate metrics. Metrics were taken from Wessel *et al.* (2008). We tested the equality of medians for each metric upstream and downstream of the three major treatment plants discharging to the CAWS. The data are not normally distributed and could not be readily transformed to approximate a normal distribution. Therefore we performed the non-parametric Kruskal-Wallis 'ANOVA' test. We concluded that, for most metrics, there was no difference between the median macroinvertebrate metrics upstream and downstream of the District's three major WRPs (Table 8).

A few metrics do, however, show a statistical difference upstream and downstream of the WRPs. In no case do the results of the Kruskal-Wallis test using ponar data agree with the results from the same test using hester-dendy data. This supports our belief that the sampling protocols measure different populations. We therefore present Table 8 with caution, and remind readers that non-parametric methods, while more robust (fewer assumptions), do not have the power of parametric methods. That said, the following conclusions can be made from the Kruskal-Wallis testing: % collector-filterers (CF) and T\_BFPOM metrics, indicate differences between upstream and downstream benthic communities at the North Side and Stickney WRPs. The T\_BFPOM metric measures the ratio of the total number of collector filterers to the total number of collector gatherers. At the Calumet WRP, the median percentage of EPT taxa and the median percentage of Tricoptera taxa from the hester-dendy samples are statistically different upstream and downstream. The percentage of Diptera (and percentage of chironomids) from the ponar samples also show significant differences upstream and downstream of the Calumet WRP.

Table 8

**P-VALUES FROM TESTS OF EQUAL MEDIANS IN THE MACROINVERTEBRATE METRICS UPSTREAM AND DOWNSTREAM OF THREE WRPS**

Metric	Reach					
	North Side WRP (Reaches 1 & 2)		Stickney WRP (Reaches 10 & 11)		Calumet WRP (Reaches 17 & 18)	
	PN	HD	PN	HD	PN	HD
% Collector-Filterer	0.0094*	0.8413	0.0013*	0.7313	0.0696	0.8734
% Collector-Gatherer	0.8894	0.0106*	0.1103	0.0721	0.9154	0.9154
C FPOM	0.5931	0.0472*	0.4810	0.0612	0.2120	0.2403
Diptera Richness	0.9776	0.9776	0.4494	0.1605	0.0748	0.1305
EPT DIP	1.0	0.6864	0.8522	0.1639	1.0	0.9570
EPT Richness	1.0	0.5717	0.8888	0.1522	1.0	0.4085
Ephemeroptera Richness	1.0	0.1464	0.2705	0.2673	1.0	0.0614
FFG DIV	0.8014	0.8673	0.6959	0.2529	0.1487	0.5538
HAB STAB	0.1082	0.1882	0.0067*	0.9267	0.0547	0.9154
% Chironomidae	0.7595	0.2433	0.9674	0.3686	0.0250*	0.75
% Diptera	0.7595	0.2433	0.8380	0.3686	0.0250*	0.75
% Dominance	0.6565	0.7389	0.1522	0.0956	0.9154	0.2882
% Ephemeroptera	1.0	0.1332	0.2705	0.2673	1.0	0.0614
% EPT	1.0	0.5184	0.8522	0.0650	1.0	0.0424*
% Oligochaeta	0.6565	0.6171	0.1522	0.1360	0.9154	0.6708
% Tricoptera	1.0	0.8250	0.3642	0.0500*	1.0	0.0424*
% Predators	1.0	0.0015*	1.0	0.1355	0.0534	0.9154
P R FFG	0.4647	0.1386	0.7964	0.4081	0.3248	1.0
Taxa Richness	0.8451	1.0	0.7600	0.1404	0.3931	0.8721
% Scrapers	0.6863	0.4401	0.9258	0.9808	0.3267	0.004*
% Shredders	0.5931	0.0883	0.4810	0.0575	0.2120	0.2002
T BFPOM	0.0076*	0.8413	0.0013*	0.7313	0.0696	0.9576
Tricoptera Richness	1.0	0.7565	0.3642	0.1445	1.0	0.8073

\*p≤0.05. Upstream and downstream reaches are statistically different.

## References

- Coffman, W. P. and L.C. Ferrington. 1996. *In An Introduction to the Aquatic Insects of North America*, Third Edition. R. W. Merrit and K.W. Cummins (*eds.*). Kendall/Hunt Publishing Company, Dubuque, IA.
- EA Engineering, Science and Technology, Inc. (EA). 2006. A Study of the Benthic Macroinvertebrate Community in Selected Metropolitan Area Waterways during 2003 and 2004. Metropolitan Water Reclamation District Report #07-47.
- Smith, D.G. 2001. *Pennak's Freshwater Invertebrates of the United States*. Fourth Edition. John Wiley & Sons, Inc. New York.
- United States Department of Agriculture (USDA). 2008. National Invasive Species Information Center. <http://www.invasivespeciesinfo.gov/aquatics/quagga.shtml>. Accessed 1/12/09.
- United States Environmental Protection Agency (USEPA). 2006. Concepts and Approaches for the Bioassessment of Non-wadeable Streams and Rivers. EPA/600/R-06/127. Washington, D.C.
- United States Geological Survey (USGS). 2008. Zebra and Quagga Mussel Resource Page. <http://nas.er.usgs.gov/taxgroup/mollusks/zebramussel/>. Accessed 01/12/09.
- Wessel, K.J., R.W. Merrit, J.O. Wilhelm, J.D.Allan, K.W. Cummins, and D.G. Uzarski. 2008. Biological evaluation of Michigan's non-wadeable rivers using macroinvertebrates. *Aquatic Ecosystem Health & Management* 11(3):335-351.